

BCRANK: predicting binding site consensus from ranked DNA sequences

Adam Ameur

June 4, 2025

1 Introduction

This document describes the **BCRANK** R package. **BCRANK**[1] is a method that takes a ranked list of genomic regions as input and outputs short DNA sequences that are overrepresented in some part of the list. The algorithm was developed for detecting transcription factor (TF) binding sites in a large number of enriched regions from high-throughput ChIP-chip or ChIP-seq experiments, but it can be applied to any ranked list of DNA sequences.

1.1 Input data

BCRANK takes a fasta file with DNA sequences as input:

- The sequences in the file must be ordered, with the ones most likely to contain a TF binding sequence at the top.
- The sequences may be of varying lengths. Usually the average length is somewhere in the range between 50bp and 2kb.
- All IUPAC nucleotide symbols are allowed in the sequences, but positions with other letters than (A, C, G, T) will not be considered in the motif search.

1.2 The algorithm

BCRANK uses a heuristic search strategy. First a score is computed for an initial short consensus sequence, typically selected at random. The score takes into account both the number of consensus occurrences and the rank of the genomic regions. Then all consensus sequences in a neighborhood of the start guess are evaluated and the one with highest score is kept as the starting point for the next iteration. When a local optimum is found, the algorithm is terminated and the locally optimal consensus is reported as a result. In order to increase the chance of detecting the globally optimal solution, the algorithm may be restarted several times using different random starting points. Alternatively, BCRANK can be used for assigning scores to previously established consensus sequences. The sections below describe in more detail how the neighborhood, scoring function and start guess are implemented.

BCRANK can extend and shorten motifs and can therefore be used in situations where the motif length is not known a priori. Moreover, BCRANK implements two optional penalties that can give relatively higher scores some consensus sequences. The effect of the penalties is that i) the consensus contains fewer redundant bases (i.e. other bases than A, C, G, T), and ii) the consensus will not be frequently occurring as a repetitive element in the enriched regions.

1.2.1 Neighborhood

In BCRANK, all consensus sequences are represented by IUPAC nucleotide symbols. The neighborhood of one consensus sequence s consists of all consensuses that can be generated from s by first adding one IUPAC letter N (representing any nucleotide) to either side of s and then flipping any base to any other IUPAC symbol. Since there are 15 symbols in total, a sequence of length l will have $14 \cdot (l + 2)$ neighbors. After each search step any flanking Ns are removed from the highest scoring sequence in the neighborhood. The removal and additions of flanking Ns allows the algorithm to shorten and extend the predicted binding sites.

1.2.2 Scoring function

The score tells whether a given consensus sequence is overrepresented in some part of the ranked list or not. Starting from N ranked regions and a consensus sequence c , a binary vector of size N is created, with 1 at position i if c is occurring in sequence number i , and 0 if not. The reverse complement of c is also allowed to match. Then the cumulative sum of the match vector is computed and stored in a vector called A . The A -vector tells where in the ranked list most occurrences are located (see Figure 1).

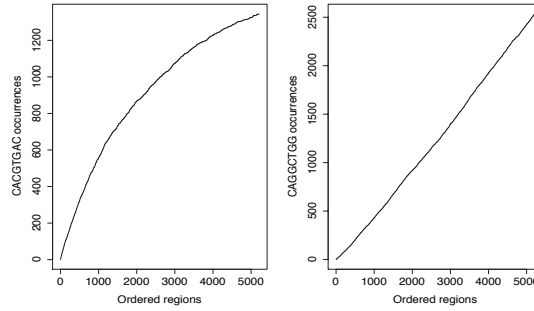


Figure 1: A -vectors for the two consensus sequences CACGTGAC (left) and CAGGCTGG (right). On the x-axis are the top 5211 regions from a whole genome ChIP-chip study on USF1 in human liver cells[2], ranked by their enrichment signal. The aim of BCRANK is to detect sequences that are biased towards some part of the list. Therefore CACGTGAC will get a higher score than CAGGCTGG even though it has a lower number of total occurrences. It is important to have enough number of ranked input regions to BCRANK, so this bias is observed for the correct binding motif. The established USF1 binding sequence is CACGTG.

To compute a score, A is compared to what it would look like if the genomic regions were randomly ordered. Therefore a large number R (the `reorderings` parameter to `bcrank()`) of random orderings of the input regions are generated, and a corresponding vector A_j is computed for each re-ordering $1 \leq j \leq R$ as above. For each j , the difference D_j between A_j and A is estimated by the area between the corresponding lines (see Figure 2). When calculating D_j , the A and A_j vectors are first scaled so they range between 0 and 1.

D_j will be close to zero when the consensus occurrences are distributed as expected by random sampling. If on the other hand all D_j are far off from zero, c is biased towards some part of the list. Therefore the score is calculated as the t-statistic T for the D_j being drawn from a distribution centered around zero. Consensus sequences that are biased towards some part of the list will thus get high scores whereas consensuses with no bias will get low scores. Moreover, consensuses that are matching just a few regions will not get a high T even if it is matching only among the top ranked regions. This is because there will be a high variation within the D_j values which will result in a low T .



Figure 2: The vector A (red line) and a corresponding vector A_j (blue line) for **CACGTGAC** in the USF1 data. There is a clear bias towards the high scoring sequences as indicated by the red line. The significance of this bias can be estimated by comparing to the **CACGTGAC**-occurrences in randomly ordered regions, as indicated by the blue line. D_j corresponds to the grey area between the two lines.

Penalties

The t-statistic gives consensus sequences that are biased towards some part of the list. But there may be other issues to take into account if the aim is to detect TFBS from ChIP-chip or ChIP-seq data. Therefore, BCRANK implements two optional penalties, $P1$ and $P2$, with values between 0 and 1. The final scoring function is defined as: $score = T \cdot P1 \cdot P2$. If a penalty is not used it will be set to 1.

- $P1$ - Penalty on non-specific bases. Let l be the length of the consensus sequence and b the total number of fixed bases (A, C, G, T) in the sequence. If there are no fixed bases, b is set to 0.5. The penalty is then defined as $P1 = b/l$.
- $P2$ - Penalty on repetitive motifs. Let $r_n, n \in 1, 2$ be the number of input DNA regions that contain at least n occurrences of the consensus. Then $P2 = 1 - (r_2/r_1)$.

1.2.3 Start guess

In case the algorithm is used for ab initio search, the initial guess is a randomly generated consensus of a specified length (the `length` parameter to `bcrank()`), with 10 bases as default. Multiple restarts with different random start guesses are usually required to increase the chance of finding the globally optimal solution. The number of restarts is determined by the `restarts` parameter.

BCRANK can also use start guesses passed to `bcrank()` by the `startguesses` parameter. By setting the `do.search` parameter to `FALSE`, BCRANK assigns scores for the given start guesses without performing any search.

1.3 Additional information

Some other important details:

- The algorithm randomly re-orders the data when the score is calculated. This implies that the same consensus sequence will get different BCRANK scores in the same data when run with different re-orderings. The variability in scores can be decreased by increasing the `reorderings` parameter
- The algorithm performs a breadth-first search, meaning that the highest scoring neighbor in the neighborhood is selected in each search step.

- The algorithm keeps track of all consensus sequences that have already been tested so the same sequence is not visited twice when performing a search.

1.4 Citation

To cite BCRANK, please use (Ameur et al, 2009), see [1] in the References section.

2 BCRANK - An example run

The user is required to load the package using the `library()` command:

```
> library(BCRANK)
```

2.1 Sequence data

BCRANK takes a fasta file containing ranked sequences as input. The command below loads an example file containing 2500 ranked regions from a whole genome ChIP-chip experiment for the protein USF1 in the human liver cell line HepG2[2].

```
> fastaFile <- system.file("Exfiles/USF1_small.fa", package="BCRANK")
```

2.2 Running BCRANK

The `bcrank()` function call below runs the BCRANK algorithm on the example USF1 data set. The `set.seed()` call sets seed for the random number generator for reproducibility.

```
> set.seed(0)
> BCRANKout <- bcrank(fastaFile, restarts=25, use.P1=TRUE, use.P2=TRUE)
```

Since it takes some time to run the algorithm, results can instead be loaded from a previous run on a larger USF1 data set containing the top 5211 regions:

```
> data(BCRANKout)
```

2.3 BCRANK output

An object of type `BCRANKresult` is returned:

```
> BCRANKout
```

An object of class "BCRANKresult"

Top 25 DNA motifs predicted by BCRANK:

	Consensus	Score
1	GTCACGTG	316.34270
2	CACGTGAC	304.59499
3	CGCGGA	147.04100
4	GCGAST	135.22207
5	AHATAATAA	128.92440
6	GCGGNGCG	121.87198
7	TNCDGGGCG	119.76454
8	GCGGGGVNG	119.65945
9	CCGCGNTBY	118.77481
10	GDGCGGHGH	115.19679
11	TNCGCGNDG	112.95972
12	CGGGNGMGC	111.93052
13	GNNDCTCGCS	111.12770
14	CGCGNNBCTBC	107.91403

```

15   CGGWSCVGA  93.88154
16   TNNCCAACG  90.63345
17   GCACANAT   84.98763
18   KTNAGABCCT  84.08165
19   TATANBDAC  80.98513
20   AACAKADTDA  79.75340
21   CCVVDDGGACG 78.86994
22   CTCRATTGHT  74.67093
23   CTATKANYWMA 68.48067
24   CBCNCGHANTR 67.87154
25   KCRTKAHTDC  52.30902

```

Use methods `toptable(object)` and `fname(object)` to access object slots.

2.3.1 The BCRANKsearch object

Use the `toptable()` function to access information about each motif found by `bcrank`. It returns an object of type `BCRANKsearch`. Here we extract the top scoring motif:

```

> topMotif <- toptable(BCRANKout,1)
> topMotif

```

An object of class "BCRANKsearch"

Search path, starting from VGKTHVBRTB:

	Consensus	Score
1	VGKTHVBRTB	6.516874
2	CGKTHVBRTB	47.627369
3	CGKTHVBRTG	73.217992
4	CGKTCVBRTG	88.236721
5	CGKTCVCRTG	105.752733
6	GKTCVCRTG	142.234278
7	GKTCACRTG	188.156513
8	GGTCACRTG	235.594317
9	GGTCACGTG	255.725078
10	GTCACGTG	316.342696

Position weight matrix for search result (GTCACGTG):

	1	2	3	4	5	6	7	8
A	0	0	0	1	0	0	0	0
C	0	0	1	0	1	0	0	0
G	1	0	0	0	0	1	0	1
T	0	1	0	0	0	0	1	0

Use methods `searchPath(object)` and `pwm(object)` to access object slots.

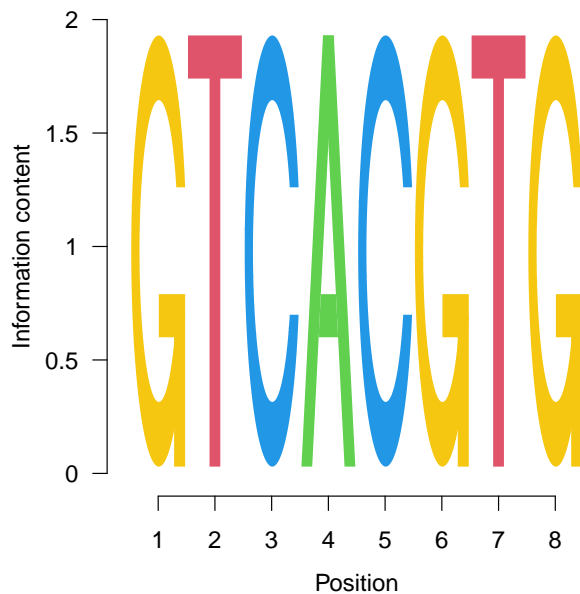
The `pwm()` function returns the position weight matrix for the search result. If `normalize` is set to `TRUE` each column will sum to 1.

```
> weightMatrix <- pwm(topMotif, normalize=FALSE)
> weightMatrix
```

	1	2	3	4	5	6	7	8
A	0	0	0	759	0	0	0	0
C	0	0	759	0	759	0	0	0
G	759	0	0	0	0	759	0	759
T	0	759	0	0	0	0	759	0

A weight matrix can be viewed as a sequence logo by using the `seqLogo` package. Make sure that the pwm is normalized before running the `seqLogo()` function.

```
> weightMatrixNormalized <- pwm(topMotif, normalize=TRUE)
> library(seqLogo)
> seqLogo(weightMatrixNormalized)
```



The search path can be visualized. For each consensus in the search path, the number of occurrences among the ranked regions are plotted. As seen in the figure, **BCRANK** searches for consensus sequences that don't give straight lines.

```
> plot(topMotif)
```



2.3.2 Predicted binding sites

Individual predicted binding sites can be reported by the `matchingSites` function.

```
> topConsensus <- as.character(toptable(BCRANKout)[1,"Consensus"])
> print(topConsensus)
```

```
[1] "GTCACGTG"
```

```
> bindingSites <- matchingSites(fastaFile,topConsensus)
> nrSites <- nrow(bindingSites)
> cat("Number predicted binding sites:",nrSites,"\n")
```

```
Number predicted binding sites: 842
```

```
> print(bindingSites[1:15,])
```

	Region header	Region nr	Start	End	Strand	Sequence
1	8:71743620-71745120:4.234	seq1	513	520	+	GTCACGTG
2	8:71743620-71745120:4.234	seq1	515	522	-	CACGTGAC
3	9:94126661-94128161:3.921	seq2	795	802	+	GTCACGTG
4	14:34660455-34661955:3.845	seq3	794	801	-	CACGTGAC
5	3:20201537-20203037:3.752	seq4	1209	1216	+	GTCACGTG
6	3:20201537-20203037:3.752	seq4	1273	1280	+	GTCACGTG
7	3:20201537-20203037:3.752	seq4	1275	1282	-	CACGTGAC
8	1:28430913-28432413:3.709	seq5	1202	1209	-	CACGTGAC
9	17:45804496-45805996:3.659	seq6	834	841	+	GTCACGTG
10	17:45804496-45805996:3.659	seq6	836	843	-	CACGTGAC
11	6:151814107-151815607:3.636	seq7	927	934	-	CACGTGAC

12	10:45541609-45543109:3.605	seq8	1036	1043	+	GTCACGTG
13	10:45541609-45543109:3.605	seq8	1051	1058	+	GTCACGTG
14	10:45541609-45543109:3.605	seq8	1038	1045	-	CACGTGAC
15	6:33493266-33494766:3.552	seq9	833	840	+	GTCACGTG

As seen in the example above, some binding sites can be reported both on the sense and anti-sense strands. If the consensus is palindromic, duplicate entries can be avoided by setting the `revComp` argument in the `matchingSites()` call to `FALSE`.

References

- [1] Ameur, A., Rada-Iglesias, A., Komorowski, J., Wadelius, C. *Identification of candidate regulatory SNPs by combination of transcription factor binding site prediction, SNP genotyping and haploChIP*. Nucleic Acids Res, 2009, 37(12):e85.
- [2] Rada-Iglesias, A., Ameur, A., Kapranov, P., Enroth, S., Komorowski, J., Gingeras, T. R., Wadelius, C. *Whole-genome maps of USF1 and USF2 binding and histone H3 acetylation reveal new aspects of promoter structure and candidate genes for common human disorders*. Genome Res, 2008, 18(3):380-92.