# Package 'scATAC.Explorer'

# December 19, 2024

**Title** A Collection of Single-cell ATAC Sequencing Datasets and Corresponding Metadata

Version 1.13.0

### **Description**

This package provides a tool to search and download a collection of publicly available single cell ATAC-seq datasets and their metadata. scATAC-Explorer aims to act as a single point of entry for users looking to study single cell ATAC-seq data. Users can quickly search available datasets using the metadata table and download datasets of interest for immediate analysis within R.

License Artistic-2.0 **Encoding UTF-8** LazyData FALSE **Roxygen** list(markdown = TRUE) RoxygenNote 7.1.1 VignetteBuilder knitr **Suggests** BiocStyle, knitr, rmarkdown, testthat (>= 3.0.0) Imports methods, Matrix **Depends** R (>= 4.1), SingleCellExperiment, BiocFileCache, data.table, utils, S4Vectors, zellkonverter biocViews SingleCellData, SequencingData, ExpressionData, GEO, Tissue, Genome, PackageTypeData BugReports https://github.com/shooshtarilab/scATACseq/issues Config/testthat/edition 3 git\_url https://git.bioconductor.org/packages/scATAC.Explorer git\_branch devel git\_last\_commit 47b6a6e git\_last\_commit\_date 2024-11-12 **Repository** Bioconductor 3.21 Date/Publication 2024-12-19

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Author Arrian Gibson-Khademi [aut, cre],
Erik Christensen [aut],
Jonathan Wang [aut],
Parisa Shooshtari [aut]
```

Maintainer Arrian Gibson-Khademi <agibsonk@uwo.ca>

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# **Description**

This function allows you to search and subset included scATAC-seq datasets. A named list of scATAC-seq\_data objects matching the provided options will be returned. Some included datasets are represented using multiple matrices. Each matrix will be a seperate named object within the list. The returned list is named by matrix allow easy identification of data. If queryATAC is called without any options it will retrieve all available datasets in sparse matrix format. This should only be done on machines with a large amount of ram (>64gb) because some datasets are quite large. In most cases it is recommended to instead filter databases with some criteria.

#### Usage

```
queryATAC(
  accession = NULL,
  author = NULL,
  journal = NULL,
 year = NULL,
  pmid = NULL,
  sequence_tech = NULL,
  score_type = NULL,
  has_cluster_annotation = NULL,
  has_cell_type_annotation = NULL,
  organism = NULL,
  genome_build = NULL,
  broad_cell_category = NULL,
  tissue_cell_type = NULL,
  disease = NULL,
 metadata_only = FALSE,
  sparse = TRUE
)
```

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# **Arguments**

accession	Search by geo accession number. Good for returning individual datasets	
author	Search by the author who published the dataset	
journal	Search by the journal the dataset was published in.	
year	Search by exact year or year ranges with '<', '>', or '-'. For example, you can return datasets newer than 2013 with '>2013'	
pmid	Search by Pubmed ID associated with the study. Good for returning individual datasets	
sequence_tech	Search by sequencing technology used to sample the cells.	
score_type has_cluster_an	Search by type of score (TPM, FPKM, raw count) notation	
	Return only those datasets that have clustering results available, or only those without (TRUE/FALSE)	
has_cell_type_annotation		
	Return only those datasets that have cell-type annotations available, or only those without annotations (TRUE/FALSE)	
organism	Search by source organism used in the study, for example human or mouse.	
genome_build	Return datasets built only using specified genome build (ex. hg19)	
broad_cell_category		
	Return datasets based on broad cell categories (ex. Hematopoetic cells). To view all cell categories available, explore the metadata table	
tissue_cell_type		
	Return datasets based on tissue or cell types sampled (ex. PBMCs, Bone marrow, Oligodendrocytes)	
disease	Return datasets based on sampled disease (ex. carcinoma, leukemia, diabetes)	
metadata only	Return rows of metadata instead of actual datasets. Useful for exploring what	

metadata\_only Return rows of metadata instead of actual datasets. Useful for exploring what

data is available without actually downloading data. Defaults to FALSE

sparse Return expression as a sparse matrix. Reccomended to use sparse format, as

dense formats tend to be excessively large.

# Value

A list containing a table of metadata or one or more SingleCellExperiment objects

# **Examples**

```
## Retrieve the metadata table to see what data is available
res <- queryATAC(metadata_only = TRUE)

## Retrieve a single dataset based on its accession number
res <- queryATAC(accession = "GSE129785")

## Retrieve the metadata of datasets between 2016 and 2020
res = queryATAC(year = "2016-2020", metadata_only = TRUE)</pre>
```

4 saveATAC

saveATAC

A function to save a scATAC-seq dataset stored in a SingleCellExperiment

# **Description**

This function allows you to save the counts, peaks, cell ID's/barcodes, and any cell clustering data to disk in csv format. It takes two options: an object to save and a directory to save in. Multiple files will be created in the provided output directory, one for each type of data available in the scATAC\_data object (counts, cell ID/Barcode, peak regions, cell type/cluster annotations).

# Usage

```
saveATAC(object, outdir, format = "mtx")
```

# **Arguments**

object The SingleCellExperiment object to be written to disk, this should be an indi-

vidual dataset returned by queryATAC.

outdir The directory to save the data in, the directory should not exist yet.

format The format to save the data in, the default is Matrix Market File Format. Alter-

native format is h5ad.

#### Value

Nothing

# **Examples**

```
# Retrieve a previously identified dataset (see queryATAC) and save it to disk
res <- queryATAC(accession = 'GSE89362')[[1]]
saveATAC(res, output_directory_name)</pre>
```

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