

# Package ‘lemur’

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**Type** Package

**Title** Latent Embedding Multivariate Regression

**Version** 1.5.0

**Description** Fit a latent embedding multivariate regression (LEMUR) model to multi-condition single-cell data. The model provides a parametric description of single-cell data measured with treatment vs. control or more complex experimental designs. The parametric model is used to (1) align conditions, (2) predict log fold changes between conditions for all cells, and (3) identify cell neighborhoods with consistent log fold changes. For those neighborhoods, a pseudobulked differential expression test is conducted to assess which genes are significantly changed.

**URL** <https://github.com/const-ae/lemur>

**BugReports** <https://github.com/const-ae/lemur/issues>

**License** MIT + file LICENSE

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---

```
.DollarNames.lemur_fit  
    Access values from a lemur_fit
```

---

### Description

Access values from a lemur\_fit

### Usage

```
## S3 method for class 'lemur_fit'  
.DollarNames(x, pattern = "")  
  
## S4 method for signature 'lemur_fit'  
x$name  
  
## S4 replacement method for signature 'lemur_fit'  
x$name <- value
```

### Arguments

x	the lemur_fit
pattern	the pattern from looking up potential values interactively
name	the name of the value behind the dollar
value	the replacement value. This only works for colData and rowData.

### Value

The respective value stored in the lemur\_fit object.

### See Also

[lemur\\_fit](#) for more documentation on the accessor functions.

---

align_harmony	<i>Enforce additional alignment of cell clusters beyond the direct differential embedding</i>
---------------	-----------------------------------------------------------------------------------------------

---

### Description

Enforce additional alignment of cell clusters beyond the direct differential embedding

**Usage**

```
align_harmony(
  fit,
  design = fit$alignment_design,
  ridge_penalty = 0.01,
  max_iter = 10,
  ...,
  verbose = TRUE
)

align_by_grouping(
  fit,
  grouping,
  design = fit$alignment_design,
  ridge_penalty = 0.01,
  preserve_position_of_NAs = FALSE,
  verbose = TRUE
)
```

**Arguments**

<code>fit</code>	a <code>lemur_fit</code> object
<code>design</code>	a specification of the design (matrix or formula) that is used for the transformation. Default: <code>fit\$design_matrix</code>
<code>ridge_penalty</code>	specification how much the flexibility of the transformation should be regularized. Default: <code>0.01</code>
<code>max_iter</code>	argument specific for <code>align_harmony</code> . The number of iterations. Default: <code>10</code>
<code>...</code>	additional parameters that are passed on to relevant functions
<code>verbose</code>	Should the method print information during the fitting. Default: <code>TRUE</code> .
<code>grouping</code>	argument specific for <code>align_by_grouping</code> . Either a vector which assigns each cell to one group or a matrix with <code>ncol(fit)</code> columns where the rows are a soft-assignment to a cluster (i.e., columns sum to 1). NA's are allowed.
<code>preserve_position_of_NAs</code>	argument specific for <code>align_by_grouping</code> . Boolean flag to decide if NAs in the grouping mean that these cells should stay where they are (if possible) or if they are free to move around. Default: <code>FALSE</code>

**Value**

The fit object with the updated `fit$embedding` and `fit$alignment_coefficients`.

**Examples**

```
data(glioblastoma_example_data)
fit <- lemur(glioblastoma_example_data, design = ~ patient_id + condition,
            n_emb = 5, verbose = FALSE)
# Creating some grouping for illustration
```

```
cell_types <- sample(c("tumor cell", "neuron", "leukocyte"), size = ncol(fit), replace = TRUE)
fit_al1 <- align_by_grouping(fit, grouping = cell_types)

# Alternatively, use harmony to automatically group cells
fit_al2 <- align_harmony(fit)
fit_al2

# The alignment coefficients are a 3D array
fit_al2$alignment_coefficients
```

---

align\_impl

*Align the points according to some grouping*

---

### Description

Align the points according to some grouping

### Usage

```
align_impl(
  embedding,
  grouping,
  design_matrix,
  ridge_penalty = 0.01,
  preserve_position_of_NAs = FALSE,
  calculate_new_embedding = TRUE
)
```

### Value

A list with the new embedding and the coefficients

---

find\_de\_neighborhoods *Find differential expression neighborhoods*

---

### Description

Find differential expression neighborhoods

**Usage**

```

find_de_neighborhoods(
  fit,
  group_by,
  contrast = fit$contrast,
  selection_procedure = c("zscore", "contrast"),
  directions = c("random", "contrast", "axis_parallel"),
  min_neighborhood_size = 50,
  de_mat = SummarizedExperiment::assays(fit)[["DE"]],
  test_data = fit$test_data,
  test_data_col_data = NULL,
  test_method = c("glmGamPoi", "edgeR", "limma", "none"),
  continuous_assay_name = fit$use_assay,
  count_assay_name = "counts",
  size_factor_method = NULL,
  design = fit$design,
  alignment_design = fit$alignment_design,
  add_diff_in_diff = TRUE,
  make_neighborhoods_consistent = FALSE,
  skip_confounded_neighborhoods = FALSE,
  control_parameters = NULL,
  verbose = TRUE
)

```

**Arguments**

<code>fit</code>	the <code>lemur_fit</code> generated by <code>lemur()</code>
<code>group_by</code>	If the <code>independent_matrix</code> is provided, <code>group_by</code> defines how the pseudobulks are formed. This is typically the variable in the column data that represents the independent unit of replication of the experiment (e.g., the mouse or patient ID). The argument has to be wrapped in <code>vars(...)</code> .
<code>contrast</code>	a specification which contrast to fit. This defaults to the <code>contrast</code> argument that was used for <code>test_de</code> and is stored in <code>fit\$contrast</code> .
<code>selection_procedure</code>	specify the algorithm that is used to select the neighborhoods for each gene. Broadly, <code>selection_procedure = "zscore"</code> is faster but less precise than <code>selection_procedure = "contrast"</code> .
<code>directions</code>	a string to define the algorithm to select the direction onto which the cells are projected before searching for the neighborhood. <code>directions = "random"</code> produces denser neighborhoods, whereas <code>directions = "contrast"</code> has usually more power. Alternatively, this can also be a matrix with one direction for each gene (i.e., a matrix of size <code>nrow(fit) * fit\$n_embedding</code> ).
<code>min_neighborhood_size</code>	the minimum number of cells per neighborhood. Default: 50.
<code>de_mat</code>	the matrix with the differential expression values and is only relevant if <code>selection_procedure = "zscore"</code> or <code>directions = "random"</code> . Defaults to an assay called "DE" that

is produced by `lemur::test_de()`.

<code>test_data</code>	a <code>SummarizedExperiment</code> object or a named list of matrices. The data is used to test if the neighborhood inferred on the training data contain a reliable significant change. If <code>test_method</code> is "glmGamPoi" or "edgeR" a test using raw counts is conducted and two matching assays are needed: (1) the continuous assay (with <code>continuous_assay_name</code> ) is projected onto the LEMUR fit to find the latent position of each cell and (2) the count assay ( <code>count_assay_name</code> ) is used for forming the pseudobulk. If <code>test_method == "limma"</code> , only the continuous assay is needed. The arguments defaults to the test data split of when calling <code>lemur()</code> .
<code>test_data_col_data</code>	additional column data for the <code>test_data</code> argument.
<code>test_method</code>	choice of test for the pseudobulked differential expression. <code>glmGamPoi</code> and <code>edgeR</code> work on an count assay. <code>limma</code> works on the continuous assay.
<code>continuous_assay_name, count_assay_name</code>	the assay or list names of <code>independent_data</code> .
<code>size_factor_method</code>	Set the procedure to calculate the size factor after pseudobulking. This argument is only relevant if <code>test_method</code> is "glmGamPoi" or "edgeR". If <code>fit</code> is subsetted, using a vector with the sequencing depth per cell ensures reasonable results. Default: NULL which means that <code>colSums(assay(fit\$test_data, count_assay_name))</code> is used.
<code>design, alignment_design</code>	the design to use for the fit. Default: <code>fit\$design</code>
<code>add_diff_in_diff</code>	a boolean to specify if the log-fold change (plus significance) of the DE in the neighborhood against the DE in the complement of the neighborhood is calculated. If TRUE, the result includes three additional columns starting with "did_" short for difference-in-difference. Default: TRUE.
<code>make_neighborhoods_consistent</code>	Include cells from outside the neighborhood if they are at least 10 times in the k-nearest neighbors of the cells inside the neighborhood. Secondly, remove cells from the neighborhood which are less than 10 times in the k-nearest neighbors of the other cells in the neighborhood. Default FALSE
<code>skip_confounded_neighborhoods</code>	Sometimes the inferred neighborhoods are not limited to a single cell state; this becomes problematic if the cells of the conditions compared in the contrast are unequally distributed between the cell states. Default: FALSE
<code>control_parameters</code>	named list with additional parameters passed to underlying functions.
<code>verbose</code>	Should the method print information during the fitting. Default: TRUE.

**Value**

a data frame with one entry per gene

name The gene name.

**neighborhood** A list column where each element is a vector with the cell names included in that neighborhood.

**n\_cells** the number of cells in the neighborhood (`lengths(neighborhood)`).

**sel\_statistic** The statistic that is maximized by the `selection_procedure`.

**pval, adj\_pval, t\_statistic, lfc** The p-value, Benjamini-Hochberg adjusted p-value (FDR), the t-statistic, and the log2 fold change of the differential expression test defined by contrast for the cells inside the neighborhood (calculated using `test_method`). Only present if `test_data` is not NULL.

**did\_pval, did\_adj\_pval, did\_lfc** The measurement if the differential expression of the cells inside the neighborhood is significantly different from the differential expression of the cells outside the neighborhood. Only present if `add_diff_in_diff = TRUE`.

### Examples

```
data(glioblastoma_example_data)
fit <- lemur(glioblastoma_example_data, design = ~ patient_id + condition,
            n_emb = 5, verbose = FALSE)
# Optional alignment
# fit <- align_harmony(fit)
fit <- test_de(fit, contrast = cond(condition = "panobinostat") - cond(condition = "ctrl"))
nei <- find_de_neighborhoods(fit, group_by = vars(patient_id))
head(nei)
```

---

fold_left	<i>Fold left over a sequence</i>
-----------	----------------------------------

---

### Description

Fold left over a sequence  
 Fold right over a sequence

### Usage

```
fold_left(init)

fold_right(init)
```

### Arguments

<code>init</code>	initial value. If not specified NULL
<code>x</code>	the sequence to iterate over
<code>FUN</code>	a function with first argument named <code>elem</code> and second argument named <code>accum</code>

### Value

The final value of `accum`.



## Examples

```
## Not run:  
# This produces ...  
fold_left(0)(1:10, \(elem, accum) accum + elem)  
# ... the same as  
sum(1:10)  
  
## End(Not run)
```

---

glioblastoma\_example\_data

*The glioblastoma\_example\_data dataset*

---

## Description

The dataset is a [SingleCellExperiment](#) object subset to 5,000 cells and 300 genes. The colData contain an entry for each cell from which patient it came and to which treatment condition it belonged ("ctrl" or "panobinostat").

## Details

The original data was collected by Zhao et al. (2021).

## Value

A [SingleCellExperiment](#) object.

## References

- Zhao, Wenting, Athanassios Dovas, Eleonora Francesca Spinazzi, Hanna Mendes Levitin, Matei Alexandru Banu, Pavan Upadhyayula, Tejaswi Sudhakar, et al. "Deconvolution of Cell Type-Specific Drug Responses in Human Tumor Tissue with Single-Cell RNA-Seq." *Genome Medicine* 13, no. 1 (December 2021): 82. <https://doi.org/10.1186/s13073-021-00894-y>.

---

grassmann\_geodesic\_regression

*Solve  $d(P, \exp_p(V * x))^2$  for  $V$*

---

## Description

Solve  $d(P, \exp_p(V * x))^2$  for  $V$

**Usage**

```
grassmann_geodesic_regression(
  coordsystems,
  design,
  base_point,
  weights = 1,
  tangent_regression = FALSE
)
```

**Value**

A three-dimensional array with the coefficients  $V$ .

---

grassmann_lm	<i>Solve <math>\ Y - \exp_p(V * x) Y\ ^2_2</math> for <math>V</math></i>
--------------	--------------------------------------------------------------------------

---

**Description**

Solve  $\|Y - \exp_p(V * x) Y\|^2_2$  for  $V$

**Usage**

```
grassmann_lm(data, design, base_point, tangent_regression = FALSE)
```

**Value**

A three-dimensional array with the coefficients  $V$ .

---

harmony_new_object	<i>Create an arbitrary Harmony object so that I can modify it later</i>
--------------------	-------------------------------------------------------------------------

---

**Description**

Create an arbitrary Harmony object so that I can modify it later

**Usage**

```
harmony_new_object()
```

**Value**

The full [harmony](#) object (R6 reference class type).

---

lemur	<i>Main function to fit the latent embedding multivariate regression (LEMUR) model</i>
-------	----------------------------------------------------------------------------------------

---

**Description**

Main function to fit the latent embedding multivariate regression (LEMUR) model

**Usage**

```
lemur(
  data,
  design = ~1,
  col_data = NULL,
  n_embedding = 15,
  linear_coefficient_estimator = c("linear", "mean", "cluster_median", "zero"),
  use_assay = "logcounts",
  test_fraction = 0.2,
  ...,
  verbose = TRUE
)
```

**Arguments**

data	a matrix with observations in the columns and features in the rows. Or a SummarizedExperiment / SingleCellExperiment object
design	a formula referring to global objects or column in the colData of data and col_data argument
col_data	an optional data frame with ncol(data) rows.
n_embedding	the dimension of the $k$ -plane that is rotated through space.
linear_coefficient_estimator	specify which estimator is used to center the conditions. "linear" runs simple regression it works well in many circumstances but can produce poor results if the composition of the cell types changes between conditions (e.g., one cell type disappears). "mean", "cluster_median" and "zero" are alternative estimators, which are each supposed to be more robust against compositional changes but cannot account for genes that change for all cells between conditions. "linear" is the default as it works best with subsequent alignment steps.
use_assay	if data is a SummarizedExperiment / SingleCellExperiment object, which assay should be used.
test_fraction	the fraction of cells that are split of before the model fit to keep an independent set of test observations. Alternatively, a logical vector of length ncol(data). Default: 20% (0.2).
...	additional parameters that are passed on to the internal function lemur_impl.
verbose	Should the method print information during the fitting. Default: TRUE.

**Value**

An object of class `lemur_fit` which extends [SingleCellExperiment](#). Accordingly, all functions that work for `sce`'s also work for `lemur_fit`'s. In addition, we give easy access to the fitted values using the dollar notation (e.g., `fit$embedding`). For details see the [lemur\\_fit](#) help page.

**References**

- Ahlmann-Eltze, C. & Huber, W. (2023). Analysis of multi-condition single-cell data with latent embedding multivariate regression. bioRxiv <https://doi.org/10.1101/2023.03.06.531268>

**See Also**

[align\\_by\\_grouping](#), [align\\_harmony](#), [test\\_de](#), [find\\_de\\_neighborhoods](#)

**Examples**

```
data(glioblastoma_example_data)
fit <- lemur(glioblastoma_example_data, design = ~ patient_id + condition, n_emb = 5)
fit
```

---

lemur_fit-class	<i>The lemur_fit class</i>
-----------------	----------------------------

---

**Description**

The `lemur_fit` class extends [SingleCellExperiment](#) and provides additional accessors to get the values of the values produced by [lemur](#).

**Usage**

```
## S4 method for signature 'lemur_fit,ANY,ANY,ANY'
x[i, j, ..., drop = TRUE]

## S4 method for signature 'lemur_fit'
design(object)
```

**Arguments**

`x, i, j, ..., drop` the `lemur_fit` object and indices for the `[]` subsetting operator  
`object` the `lemur_fit` object for the [BiocGenerics::design](#) generic

**Details**

To access the values produced by `lemur`, use the dollar notation (`$`):

`fit$n_embedding` the number of embedding dimensions.

`fit$design` the specification of the design in `lemur`. Usually this is a `stats::formula`.

`fit$base_point` a matrix (`nrow(fit) * fit$n_embedding`) with the base point for the Grassmann exponential map.

`fit$coefficients` a three-dimensional tensor (`nrow(fit) * fit$n_embedding * ncol(fit$design_matrix)`) with the coefficients for the exponential map.

`fit$embedding` a matrix (`fit$n_embedding * ncol(fit)`) with the low dimensional position for each cell.

`fit$design_matrix` a matrix with covariates for each cell (`ncol(fit) * ncol(fit$design_matrix)`).

`fit$linear_coefficients` a matrix (`nrow(fit) * ncol(fit$design_matrix)`) with the coefficients for the linear regression.

`fit$alignment_coefficients` a 3D tensor with the coefficients for the alignment (`fit$n_embedding * fit$n_embedding * ncol(fit$design_matrix)`)

`fit$alignment_design` an alternative design specification for the alignment. This is typically a `stats::formula`.

`fit$alignment_design_matrix` an alternative design matrix specification for the alignment.

`fit$contrast` a parsed version of the contrast specification from the `test_de` function or `NULL`.

`fit$colData` the column annotation `DataFrame`.

`fit$rowData` the row annotation `DataFrame`.

**Value**

An object of class `lemur_fit`.

**See Also**

[lemur](#), [predict](#), [residuals](#)

**Examples**

```
# The easiest way to make a lemur_fit object, is to call `lemur`
data(glioblastoma_example_data)
fit <- lemur(glioblastoma_example_data, design = ~ patient_id + condition,
            n_emb = 5, verbose = FALSE)
```

```
fit$n_embedding
fit$embedding[,1:10]
fit$n_embedding
fit$embedding[,1:10]
fit$design_matrix[1:10,]
fit$coefficients[1:3,,]
```

---

mply_dbl	<i>Iterating function that returns a matrix</i>
----------	-------------------------------------------------

---

**Description**

The length of `x` determines the number of rows. The length of `FUN(x[i])` determines the number of columns. Must match `ncol`.

**Usage**

```
mply_dbl(x, FUN, ncol = 1, ...)
```

```
stack_rows(x)
```

```
stack_cols(x)
```

**Arguments**

<code>x</code>	the sequence that is mapped to a matrix
<code>FUN</code>	the function that returns a vector of length <code>ncol</code>
<code>ncol</code>	the length of the output vector
<code>...</code>	additional arguments that are passed to <code>FUN</code>

**Value**

A matrix with `length(x) / nrow(x)` rows and `ncol` columns. For `mply_dbl` the number of columns depends on the output of `FUN`.

**Functions**

- `stack_rows()`: Each list element becomes a row in a matrix
- `stack_cols()`: Each list element becomes a row in a matrix

---

one_hot_encoding	<i>Take a vector and convert it to a one-hot encoded matrix</i>
------------------	-----------------------------------------------------------------

---

**Description**

Take a vector and convert it to a one-hot encoded matrix

**Usage**

```
one_hot_encoding(groups)
```

**Value**

A matrix with `length(unique(groups))` rows and `length(groups)` columns.

---

predict.lemur\_fit      *Predict values from lemur\_fit object*

---

## Description

Predict values from lemur\_fit object

## Usage

```
## S3 method for class 'lemur_fit'
predict(
  object,
  newdata = NULL,
  newdesign = NULL,
  newcondition = NULL,
  embedding = object$embedding,
  with_linear_model = TRUE,
  with_embedding = TRUE,
  with_alignment = TRUE,
  ...
)
```

## Arguments

object	an lemur_fit object
newdata	a data.frame which passed to <code>model.matrix</code> with design to make the newdesign matrix
newdesign	a matrix with the covariates for which the output is predicted. If NULL, the <code>object\$design_matrix</code> is used. If it is a vector it is repeated <code>ncol(embedding)</code> times to create a design matrix with the same entry for each cell.
newcondition	an unquoted expression with a call to <code>cond()</code> specifying the covariates of the prediction. See the contrast argument in <code>test_de</code> for more details. Note that combinations of multiple calls to <code>cond()</code> are not allowed (e.g., <code>cond(a = 1) - cond(a = 2)</code> ). If specified, <code>newdata</code> and <code>newdesign</code> are ignored.
embedding	the low-dimensional cell position for which the output is predicted.
with_linear_model	a boolean to indicate if the linear regression offset is included in the prediction.
with_embedding	a boolean to indicate if the embedding contributes to the output.
with_alignment	a boolean to indicate if the alignment effect is removed from the output.
...	additional parameters passed to <code>predict_impl</code> .

## Value

A matrix with the same dimension `nrow(object) * nrow(newdesign)`.

**See Also**[residuals](#)**Examples**

```

data(glioblastoma_example_data)
fit <- lemur(glioblastoma_example_data, design = ~ patient_id + condition,
            n_emb = 5, verbose = FALSE)

pred <- predict(fit)

pred_ctrl <- predict(fit, newdesign = c(1, 0, 0, 0, 0, 0))
pred_trt <- predict(fit, newdesign = c(1, 0, 0, 0, 0, 1))
# This is the same as the test_de result
fit <- test_de(fit, cond(condition = "panobinostat") - cond(condition = "ctrl"))
all.equal(SummarizedExperiment::assay(fit, "DE"), pred_trt - pred_ctrl,
          check.attributes = FALSE)

```

---

project\_on\_lemur\_fit *Project new data onto the latent spaces of an existing lemur fit*

---

**Description**

Project new data onto the latent spaces of an existing lemur fit

**Usage**

```

project_on_lemur_fit(
  fit,
  data,
  col_data = NULL,
  use_assay = "logcounts",
  design = fit$design,
  alignment_design = fit$alignment_design,
  return = c("matrix", "lemur_fit")
)

```

**Arguments**

fit	an lemur_fit object
data	a matrix with observations in the columns and features in the rows. Or a SummarizedExperiment / SingleCellExperiment object. The features must match the features in fit.
col_data	col_data an optional data frame with ncol(data) rows.
use_assay	if data is a SummarizedExperiment / SingleCellExperiment object, which assay should be used.



`design, alignment_design` the design formulas or design matrices that are used to project the data on the correct latent subspace. Both default to the designs from the `fit` object.

`return` which data structure is returned.

**Value**

Either a matrix with the low-dimensional embeddings of the data or an object of class `lemur_fit` wrapping that embedding.

**Examples**

```
data(glioblastoma_example_data)

subset1 <- glioblastoma_example_data[,1:2500]
subset2 <- glioblastoma_example_data[,2501:5000]

fit <- lemur(subset1, design = ~ condition, n_emb = 5,
             test_fraction = 0, verbose = FALSE)

# Returns a `lemur_fit` object with the projection of `subset2`
fit2 <- project_on_lemur_fit(fit, subset2, return = "lemur_fit")
fit2
```

---

pseudoinverse

*Moore-Penrose pseudoinverse calculated via SVD*

---

**Description**

In the simplest case, the pseudoinverse is

$$X^+ = (X^T X)^{-1} X^T.$$

**Usage**

```
pseudoinverse(X)
```

**Arguments**

`X` a matrix `X`

**Details**

To handle the more general case, the pseudoinverse can be expressed using a SVD  $X = UDV^T$ :

$$X^+ = VD^{-1}U^T$$

**Value**

The matrix  $X^+$ .

---

recursive\_least\_squares

*Iteratively calculate the least squares solution*

---

**Description**

Both functions are for testing purposes. There is a faster implementation called `cum_brls_which_abs_max`.

**Usage**

```
recursive_least_squares(y, X)

bulked_recursive_least_squares_contrast(
  y,
  X,
  group,
  contrast,
  ridge_penalty = 1e-06
)
```

**Arguments**

`y` a vector with observations  
`X` a design matrix

**Value**

a matrix where column `i` is the solution to  $y[1:i] \sim X[1:i, ]$ .

---

reexports

*Objects exported from other packages*

---

**Description**

These objects are imported from other packages. Follow the links below to see their documentation.

**glmGamPoi** [vars](#)

**Value**

see [glmGamPoi::vars](#).

**Examples**

```
# `vars` quotes expressions (just like in dplyr)
vars(condition, sample)
```

---

```
residuals,lemur_fit-method
```

*Predict values from lemur\_fit object*

---

**Description**

Predict values from lemur\_fit object

**Usage**

```
## S4 method for signature 'lemur_fit'
residuals(object, with_linear_model = TRUE, with_embedding = TRUE, ...)
```

**Arguments**

object            an lemur\_fit object  
with\_linear\_model            a boolean to indicate if the linear regression offset is included in the prediction.  
with\_embedding    a boolean to indicate if the embedding contributes to the output.  
...                ignored.

**Value**

A matrix with the same dimension `dim(object)`.

**See Also**

[predict.lemur\\_fit](#)

**Examples**

```
data(glioblastoma_example_data)
fit <- lemur(glioblastoma_example_data, design = ~ patient_id + condition,
            n_emb = 5, verbose = FALSE)

resid <- residuals(fit)
dim(resid)
```

---

ridge_regression	<i>Ridge regression</i>
------------------	-------------------------

---

**Description**

The function does not treat the intercept special.

**Usage**

```
ridge_regression(Y, X, ridge_penalty = 0, weights = rep(1, nrow(X)))
```

**Arguments**

Y	the observations matrix (features x samples)
X	the design matrix (samples x covariates)
ridge_penalty	a numeric vector or matrix of size (covariates or covariates x covariates respectively)
weights	a vector of observation weights

**Value**

The matrix of coefficients.

---

stack_slice	<i>Make a cube from a list of matrices</i>
-------------	--------------------------------------------

---

**Description**

The length of the list will become the third dimension of the cube.

**Usage**

```
stack_slice(x)
destack_slice(x)
```

**Arguments**

x	a list of vectors/matrices that are stacked
---	---------------------------------------------

**Value**

A three-dimensional array.

**Functions**

- `destack_slice()`: Make a list of matrices from a cube

---

test_de	<i>Predict log fold changes between conditions for each cell</i>
---------	------------------------------------------------------------------

---

**Description**

Predict log fold changes between conditions for each cell

**Usage**

```
test_de(
  fit,
  contrast,
  embedding = NULL,
  consider = c("embedding+linear", "embedding", "linear"),
  new_assay_name = "DE"
)
```

**Arguments**

fit	the result of calling <code>lemur()</code>
contrast	Specification of the contrast: a call to <code>cond()</code> specifying a full observation (e.g. <code>cond(treatment = "A", sex = "male") - cond(treatment = "C", sex = "male")</code> ) to compare treatment A vs C for male observations). Unspecified factors default to the reference level.
embedding	matrix of size $n_{\text{embedding}} \times n$ that specifies where in the latent space the differential expression is tested. It defaults to the position of all cells from the original fit.
consider	specify which part of the model are considered for the differential expression test.
new_assay_name	the name of the assay added to the fit object. Default: "DE".

**Value**

If `is.null(embedding)` the fit object with a new assay called "DE". Otherwise return a matrix with the differential expression values.

**See Also**

[find\\_de\\_neighborhoods](#)

**Examples**

```
library(SummarizedExperiment)
library(SingleCellExperiment)

data(glioblastoma_example_data)
fit <- lemur(glioblastoma_example_data, design = ~ patient_id + condition,
```

```

        n_emb = 5, verbose = FALSE)
# Optional alignment
# fit <- align_harmony(fit)
fit <- test_de(fit, contrast = cond(condition = "panobinostat") - cond(condition = "ctrl"))

# The fit object contains a new assay called "DE"
assayNames(fit)

# The DE assay captures differences between conditions
is_ctrl_cond <- fit$colData$condition == "ctrl"
mean(logcounts(fit)[1,!is_ctrl_cond]) - mean(logcounts(fit)[1,is_ctrl_cond])
mean(assay(fit, "DE")[1,])

```

---

test\_global

*Differential embedding for each condition*


---

## Description

Differential embedding for each condition

## Usage

```

test_global(
  fit,
  contrast,
  reduced_design = NULL,
  consider = c("embedding+linear", "embedding", "linear"),
  variance_est = c("analytical", "resampling", "none"),
  verbose = TRUE,
  ...
)

```

## Arguments

fit	the result of calling <code>lemur()</code>
contrast	Specification of the contrast: a call to <code>cond()</code> specifying a full observation (e.g. <code>cond(treatment = "A", sex = "male") - cond(treatment = "C", sex = "male")</code> ) to compare treatment A vs C for male observations). Unspecified factors default to the reference level.
reduced_design	an alternative specification of the null hypothesis.
consider	specify which part of the model are considered for the differential expression test.
variance_est	How or if the variance should be estimated. 'analytical' is only compatible with <code>consider = "linear"</code> . 'resampling' is the most flexible (to adapt the number of resampling iterations, set <code>n_resampling_iter</code> . Default: 100)
verbose	should the method print information during the fitting. Default: TRUE.
...	additional arguments.

**Value**

a data.frame

---

%zero\_dom\_mat\_mult%    *Helper function that makes sure that  $NA * 0 = 0$  in matrix multiply*

---

**Description**

Helper function that makes sure that  $NA * 0 = 0$  in matrix multiply

**Usage**

X %zero\_dom\_mat\_mult% Y

**Arguments**

X                    a matrix of size n\*m  
Y                    a matrix of size m\*p

**Value**

a matrix of size n\*p

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