

Package ‘TPP2D’

December 19, 2024

Title Detection of ligand-protein interactions from 2D thermal profiles (DLPTP)

Version 1.23.0

Description Detection of ligand-protein interactions from 2D thermal profiles (DLPTP), Performs an FDR-controlled analysis of 2D-TPP experiments by functional analysis of dose-response curves across temperatures.

License GPL-3

Encoding UTF-8

VignetteBuilder knitr

LazyData false

biocViews Software, Proteomics, DataImport

BugReports <https://support.bioconductor.org/>

URL <http://bioconductor.org/packages/TPP2D>

RoxygenNote 7.1.2

Depends R (>= 3.6.0), stats, utils, dplyr, methods

Imports ggplot2, tidyr, foreach, doParallel, openxlsx, stringr, RCurl, parallel, MASS, BiocParallel, limma

Suggests knitr, testthat, rmarkdown, BiocStyle

git_url <https://git.bioconductor.org/packages/TPP2D>

git_branch devel

git_last_commit 9179fb5

git_last_commit_date 2024-10-29

Repository Bioconductor 3.21

Date/Publication 2024-12-18

Author Nils Kurzawa [aut, cre],
Holger Franken [aut],
Simon Anders [aut],
Wolfgang Huber [aut],
Mikhail M. Savitski [aut]

Maintainer Nils Kurzawa <nilskurzawa@gmail.com>

Contents

annotateDataList	3
bootstrapNull	4
bootstrapNullAlternativeModel	5
bootstrapNullAlternativeModelFast	7
competeModels	9
computeFdr-defunct	10
computeFstat	10
computeFStatFromParams	11
configWide2Long	12
config_tab	12
filterOutContaminants	13
findHits	13
fitAndEvalDataset	14
fitH0Model	15
fitH1Model	16
getFDR	17
getModelParamsDf	18
getPEC504Temperature	19
getPvalues	20
gg_qq	21
import2dDataset	22
import2dMain	23
plot2dTppFcHeatmap	25
plot2dTppFit	25
plot2dTppProfile	27
plot2dTppRelProfile	27
plot2dTppVolcano	28
raw_dat_list	29
recomputeSignalFromRatios	29
renameColumns	30
resolveAmbiguousProteinNames	31
runTPP2D	32
simulated_cell_extract_df	34
TPP2D-defunct	35
tpp2dExperiment-class	35
TPP_importCheckConfigTable	36
Index	38

annotateDataList	<i>Annotate imported data list using a config table</i>
------------------	---

Description

Annotate imported data list using a config table

Usage

```
annotateDataList(dataList, geneNameVar, configLong, intensityStr, fcStr)
```

Arguments

dataList	list of datasets from different MS runs corresponding to a 2D-TPP dataset
geneNameVar	character string of the column name that describes the gene name of a given protein in the raw data files
configLong	long formatted data frame of a corresponding config table
intensityStr	character string indicating which columns contain raw intensities measurements
fcStr	character string indicating which columns contain the actual fold change values. Those column names containing the suffix fcStr will be regarded as containing fold change values.

Value

data frame containing all data annotated by information supplied in the config table

Examples

```
data("config_tab")
data("raw_dat_list")
datalist <- import2dMain(configTable = config_tab,
                        data = raw_dat_list,
                        idVar = "protein_id",
                        fcStr = "rel_fc_",
                        addCol = "gene_name",
                        naStrs = NA,
                        intensityStr = "signal_sum_",
                        nonZeroCols = "qum",
                        qualColName = "qum")
configLong <- configWide2Long(configWide = config_tab)
annotateDataList(dataList = datalist,
                 geneNameVar = "gene_name",
                 configLong = configLong,
                 intensityStr = "signal_sum_",
                 fcStr = "rel_fc_")
```

bootstrapNull

*Bootstrap null distribution of F statistics for FDR estimation***Description**

Bootstrap null distribution of F statistics for FDR estimation

Usage

```
bootstrapNull(
  df,
  maxit = 500,
  independentFiltering = FALSE,
  fcThres = 1.5,
  minObs = 20,
  optim_fun_h0 = .min_RSS_h0,
  optim_fun_h1 = .min_RSS_h1_slope_pEC50,
  optim_fun_h1_2 = NULL,
  gr_fun_h0 = NULL,
  gr_fun_h1 = NULL,
  gr_fun_h1_2 = NULL,
  ncores = 1,
  B = 20,
  byMsExp = TRUE
)
```

Arguments

df	tidy data_frame retrieved after import of a 2D-TPP dataset, potential filtering and addition of a column "nObs" containing the number of observations per protein
maxit	maximal number of iterations the optimization should be given, default is set to 500
independentFiltering	boolean flag indicating whether independent filtering should be performed based on minimal fold changes per protein profile
fcThres	numeric value of minimal fold change (or inverse fold change) a protein has to show to be kept upon independent filtering
minObs	numeric value of minimal number of observations that should be required per protein
optim_fun_h0	optimization function that should be used for fitting the H0 model
optim_fun_h1	optimization function that should be used for fitting the H1 model
optim_fun_h1_2	optional additional optimization function that will be run with parameters retrieved from optim_fun_h1 and should be used for fitting the H1 model with the trimmed sum model, default is NULL

gr_fun_h0	optional gradient function for optim_fun_h0, default is NULL
gr_fun_h1	optional gradient function for optim_fun_h1, default is NULL
gr_fun_h1_2	optional gradient function for optim_fun_h1_2, default is NULL
ncores	numeric value of numbers of cores that the function should use to parallelize
B	numeric value of rounds of bootstrap, default: 20
byMsExp	boolean flag indicating whether resampling of residuals should be performed separately for data generated by different MS experiments, default TRUE, recommended

Value

data frame containing F statistics of proteins with permuted 2D thermal profiles that are informative on the Null distribution of F statistics

Examples

```
data("simulated_cell_extract_df")
temp_df <- simulated_cell_extract_df %>%
  filter(clustername %in% paste0("protein", 1:3)) %>%
  group_by(representative) %>%
  mutate(nObs = n()) %>%
  ungroup
boot_df <- bootstrapNull(temp_df, B = 2/10)
```

bootstrapNullAlternativeModel

Bootstrap null distribution of F statistics for FDR estimation based on resampling alternative model residuals

Description

Bootstrap null distribution of F statistics for FDR estimation based on resampling alternative model residuals

Usage

```
bootstrapNullAlternativeModel(
  df,
  params_df,
  maxit = 500,
  independentFiltering = FALSE,
  fcThres = 1.5,
  minObs = 20,
  optim_fun_h0 = TPP2D:::min_RSS_h0,
  optim_fun_h1 = TPP2D:::min_RSS_h1_slope_pEC50,
```

```

optim_fun_h1_2 = NULL,
gr_fun_h0 = NULL,
gr_fun_h1 = NULL,
gr_fun_h1_2 = NULL,
BPPARAM = BiocParallel::SerialParam(progressbar = TRUE),
B = 20,
byMsExp = TRUE,
verbose = FALSE
)

```

Arguments

<code>df</code>	tidy data frame retrieved after import of a 2D-TPP dataset, potential filtering and addition of a column "nObs" containing the number of observations per protein
<code>params_df</code>	data frame listing all null and alternative model parameters as obtained by 'get-ModelParamsDf'
<code>maxit</code>	maximal number of iterations the optimization should be given, default is set to 500
<code>independentFiltering</code>	boolean flag indicating whether independent filtering should be performed based on minimal fold changes per protein profile
<code>fcThres</code>	numeric value of minimal fold change (or inverse fold change) a protein has to show to be kept upon independent filtering
<code>minObs</code>	numeric value of minimal number of observations that should be required per protein
<code>optim_fun_h0</code>	optimization function that should be used for fitting the H0 model
<code>optim_fun_h1</code>	optimization function that should be used for fitting the H1 model
<code>optim_fun_h1_2</code>	optional additional optimization function that will be run with parameters retrieved from <code>optim_fun_h1</code> and should be used for fitting the H1 model with the trimmed sum model, default is NULL
<code>gr_fun_h0</code>	optional gradient function for <code>optim_fun_h0</code> , default is NULL
<code>gr_fun_h1</code>	optional gradient function for <code>optim_fun_h1</code> , default is NULL
<code>gr_fun_h1_2</code>	optional gradient function for <code>optim_fun_h1_2</code> , default is NULL
<code>BPPARAM</code>	BiocParallel parameter for optional parallelization of null distribution generation through bootstrapping, default: <code>BiocParallel::SerialParam()</code>
<code>B</code>	numeric value of rounds of bootstrap, default: 20
<code>byMsExp</code>	boolean flag indicating whether resampling of residuals should be performed separately for data generated by different MS experiments, default TRUE, recommended
<code>verbose</code>	logical indicating whether to print each protein while its profile is bootstrapped

Value

data frame containing F statistics of proteins with permuted 2D thermal profiles that are informative on the Null distribution of F statistics

Examples

```

data("simulated_cell_extract_df")
temp_df <- simulated_cell_extract_df %>%
  filter(clustername %in% paste0("protein", 1:3)) %>%
  group_by(representative) %>%
  mutate(nObs = n()) %>%
  ungroup
temp_params_df <- getModelParamsDf(temp_df)
boot_df <- bootstrapNullAlternativeModel(
  temp_df, params_df = temp_params_df, B = 2)

```

bootstrapNullAlternativeModelFast

Bootstrap null distribution of F statistics for FDR estimation based on resampling alternative model residuals with only one round of model fitting on resampled data and subsequent resampling of thereby obtained residuals

Description

Bootstrap null distribution of F statistics for FDR estimation based on resampling alternative model residuals with only one round of model fitting on resampled data and subsequent resampling of thereby obtained residuals

Usage

```

bootstrapNullAlternativeModelFast(
  df,
  params_df,
  maxit = 500,
  independentFiltering = FALSE,
  fcThres = 1.5,
  minObs = 20,
  optim_fun_h0 = TPP2D:::min_RSS_h0,
  optim_fun_h1 = TPP2D:::min_RSS_h1_slope_pEC50,
  optim_fun_h1_2 = NULL,
  gr_fun_h0 = NULL,
  gr_fun_h1 = NULL,
  gr_fun_h1_2 = NULL,
  BPPARAM = BiocParallel::SerialParam(progressbar = TRUE),
  B = 20,
  byMsExp = TRUE,
  verbose = FALSE
)

```

Arguments

<code>df</code>	tidy data frame retrieved after import of a 2D-TPP dataset, potential filtering and addition of a column "nObs" containing the number of observations per protein
<code>params_df</code>	data frame listing all null and alternative model parameters as obtained by 'get-ModelParamsDf'
<code>maxit</code>	maximal number of iterations the optimization should be given, default is set to 500
<code>independentFiltering</code>	boolean flag indicating whether independent filtering should be performed based on minimal fold changes per protein profile
<code>fcThres</code>	numeric value of minimal fold change (or inverse fold change) a protein has to show to be kept upon independent filtering
<code>minObs</code>	numeric value of minimal number of observations that should be required per protein
<code>optim_fun_h0</code>	optimization function that should be used for fitting the H0 model
<code>optim_fun_h1</code>	optimization function that should be used for fitting the H1 model
<code>optim_fun_h1_2</code>	optional additional optimization function that will be run with parameters retrieved from <code>optim_fun_h1</code> and should be used for fitting the H1 model with the trimmed sum model, default is NULL
<code>gr_fun_h0</code>	optional gradient function for <code>optim_fun_h0</code> , default is NULL
<code>gr_fun_h1</code>	optional gradient function for <code>optim_fun_h1</code> , default is NULL
<code>gr_fun_h1_2</code>	optional gradient function for <code>optim_fun_h1_2</code> , default is NULL
<code>BPPARAM</code>	BiocParallel parameter for optional parallelization of null distribution generation through bootstrapping, default: <code>BiocParallel::SerialParam()</code>
<code>B</code>	numeric value of rounds of bootstrap, default: 20
<code>byMsExp</code>	boolean flag indicating whether resampling of residuals should be performed separately for data generated by different MS experiments, default TRUE, recommended
<code>verbose</code>	logical indicating whether to print each protein while its profile is bootstrapped

Value

data frame containing F statistics of proteins with permuted 2D thermal profiles that are informative on the Null distribution of F statistics

Examples

```
data("simulated_cell_extract_df")
temp_df <- simulated_cell_extract_df %>%
  filter(clustername %in% paste0("protein", 1:3)) %>%
  group_by(representative) %>%
  mutate(nObs = n()) %>%
  ungroup
temp_params_df <- getModelParamsDf(temp_df)
boot_df <- bootstrapNullAlternativeModelFast(
  temp_df, params_df = temp_params_df, B = 20)
```

 competeModels

Compete H0 and H1 models per protein and obtain F statistic

Description

Compete H0 and H1 models per protein and obtain F statistic

Usage

```

competeModels(
  df,
  fcThres = 1.5,
  independentFiltering = FALSE,
  minObs = 20,
  optim_fun_h0 = .min_RSS_h0,
  optim_fun_h1 = .min_RSS_h1_slope_pEC50,
  optim_fun_h1_2 = NULL,
  gr_fun_h0 = NULL,
  gr_fun_h1 = NULL,
  gr_fun_h1_2 = NULL,
  maxit = 750
)

```

Arguments

df	tidy data frame retrieved after import of a 2D-TPP dataset, potential filtering and addition of a column "nObs" containing the number of observations per protein
fcThres	numeric value of minimal fold change (or inverse fold change) a protein has to show to be kept upon independent filtering
independentFiltering	boolean flag indicating whether independent filtering should be performed based on minimal fold changes per protein profile
minObs	numeric value of minimal number of observations that should be required per protein
optim_fun_h0	optimization function that should be used for fitting the H0 model
optim_fun_h1	optimization function that should be used for fitting the H1 model
optim_fun_h1_2	optional additional optimization function that will be run with parameters retrieved from optim_fun_h1 and should be used for fitting the H1 model with the trimmed sum model, default is NULL
gr_fun_h0	optional gradient function for optim_fun_h0, default is NULL
gr_fun_h1	optional gradient function for optim_fun_h1, default is NULL
gr_fun_h1_2	optional gradient function for optim_fun_h1_2, default is NULL
maxit	maximal number of iterations the optimization should be given, default is set to 500

Value

data frame summarising the fit characteristics of H0 and H1 models and thereof resulting computed F statistics per protein

Examples

```
data("simulated_cell_extract_df")
temp_df <- simulated_cell_extract_df %>%
  filter(clustername %in% paste0("protein", 1:10)) %>%
  group_by(representative) %>%
  mutate(nObs = n()) %>%
  ungroup
competeModels(temp_df)
```

computeFdr-defunct	<i>Compute FDR for given F statistics based on true and null dataset (old function)</i>
--------------------	---

Description

Compute FDR for given F statistics based on true and null dataset (old function)

See Also

[TPP2D-defunct](#)

computeFstat	<i>Compute F statistic from H1 and H0 model characteristics</i>
--------------	---

Description

Compute F statistic from H1 and H0 model characteristics

Usage

```
computeFstat(h0_df, h1_df)
```

Arguments

h0_df	data frame with H0 model characteristics for each protein
h1_df	data frame with H1 model characteristics for each protein

Value

data frame with H0 and H1 model characteristics for each protein and respectively computed F statistics

Examples

```
data("simulated_cell_extract_df")
temp_df <- simulated_cell_extract_df %>%
  filter(clustertype %in% paste0("protein", 1:20)) %>%
  group_by(representative) %>%
  mutate(nObs = n()) %>%
  ungroup

h0_df <- fitH0Model(temp_df)
h1_df <- fitH1Model(temp_df)

computeFstat(h0_df, h1_df)
```

computeFStatFromParams

Compute F statistics from parameter data frame

Description

Compute F statistics from parameter data frame

Usage

```
computeFStatFromParams(params_df)
```

Arguments

params_df data frame listing all null and alternative model parameters as obtained by 'getModelParamsDf'

Value

data frame of all proteins and computed F statistics and parameters that were used for the computation

Examples

```
data("simulated_cell_extract_df")
params_df <- getModelParamsDf(simulated_cell_extract_df)
computeFStatFromParams(params_df)
```

configWide2Long	<i>Transform configuration table from wide to long</i>
-----------------	--

Description

Transform configuration table from wide to long

Usage

```
configWide2Long(configWide)
```

Arguments

configWide data frame containing a config table

Value

data frame containing config table in long format

Examples

```
data("config_tab")
configWide2Long(configWide = config_tab)
```

config_tab	<i>Example config table for a import of a simulated 2D-TPP cell extract dataset</i>
------------	---

Description

Config table for import of simulated example dataset obtained by 2D-TPP experiments for analysis by the TPP2D-package. It's a data frame with the columns "Compound" describing the compound used for the assay, "Experiment" listing MS experiment ids of the separate runs (typically comprising two multiplexed adjacent temperature), "Temperature": the temperature used for a given sub-experiment, the respective TMT labels "126"-"131L", RefCol referring to the label used as a reference label for computing relative fold changes (usually the label used for the control treatment). Please note that when the data is not supplied as a list of already imported data frames the config table for the import function should be a path to a txt, csv or xlsx file containing an additional column "Path" listing for each row the respective path to a searched protein output file.

Usage

```
data("config_tab")
```

Format

"Compound" describing the compound used for the assay, "Experiment" listing MS experiment ids of the separate runs (typically comprising two multiplexed adjacent temperature), "Temperature": the temperature used for a given sub-experiment, the respective TMT labels "126"- "131L", RefCol referring to the label used as a reference label for computing relative fold changes (usually the label used for the control treatment).

filterOutContaminants *Filter out contaminants*

Description

Filter out contaminants

Usage

```
filterOutContaminants(dataLong)
```

Arguments

dataLong long format data frame of imported dataset

Value

data frame containing full dataset filtered to contain no contaminants

Examples

```
data("simulated_cell_extract_df")
filterOutContaminants(simulated_cell_extract_df)
```

findHits *Find hits according to FDR threshold*

Description

Find hits according to FDR threshold

Usage

```
findHits(fdr_df, alpha)
```

Arguments

fdr_df data frame obtained from computeFdr
alpha significance threshold, default is set to 0.1

Value

data frame of significant hits at $FDR \leq \alpha$

Examples

```
data("simulated_cell_extract_df")
temp_df <- simulated_cell_extract_df %>%
  filter(clustername %in% paste0("protein", 1:5)) %>%
  group_by(representative) %>%
  mutate(nObs = n()) %>%
  ungroup
example_out <- fitAndEvalDataset(temp_df)
example_null <- bootstrapNull(temp_df, B = 1)
fdr_df <- getFDR(example_out, example_null)
findHits(fdr_df, 0.1)
```

fitAndEvalDataset	<i>Fit H0 and H1 model to 2D thermal profiles of proteins and compute F statistic</i>
-------------------	---

Description

Fit H0 and H1 model to 2D thermal profiles of proteins and compute F statistic

Usage

```
fitAndEvalDataset(
  df,
  maxit = 500,
  optim_fun_h0 = .min_RSS_h0,
  optim_fun_h1 = .min_RSS_h1_slope_pEC50,
  optim_fun_h1_2 = NULL,
  gr_fun_h0 = NULL,
  gr_fun_h1 = NULL,
  gr_fun_h1_2 = NULL,
  ec50_lower_limit = NULL,
  ec50_upper_limit = NULL,
  slopEC50 = TRUE
)
```

Arguments

df	tidy data_frame retrieved after import of a 2D-TPP dataset, potential filtering and addition of a column "nObs" containing the number of observations per protein
maxit	maximal number of iterations the optimization should be given, default is set to 500

optim_fun_h0	optimization function that should be used for fitting the H0 model
optim_fun_h1	optimization function that should be used for fitting the H1 model
optim_fun_h1_2	optional additional optimization function that will be run with parameters retrieved from optim_fun_h1 and should be used for fitting the H1 model with the trimmed sum model, default is NULL
gr_fun_h0	optional gradient function for optim_fun_h0, default is NULL
gr_fun_h1	optional gradient function for optim_fun_h1, default is NULL
gr_fun_h1_2	optional gradient function for optim_fun_h1_2, default is NULL
ec50_lower_limit	lower limit of ec50 parameter
ec50_upper_limit	upper limit of ec50 parameter
slopEC50	logical flag indicating whether the h1 model is fitted with a linear model describing the shift of the pEC50 over temperatures

Value

data frame with H0 and H1 model characteristics for each protein and respectively computed F statistics

Examples

```
data("simulated_cell_extract_df")
temp_df <- simulated_cell_extract_df %>%
  group_by(representative) %>%
  mutate(nObs = n()) %>%
  ungroup
fitAndEvalDataset(temp_df)
```

fitH0Model

Fit H0 model and evaluate fit statistics

Description

Fit H0 model and evaluate fit statistics

Usage

```
fitH0Model(df, maxit = 500, optim_fun = .min_RSS_h0, gr_fun = NULL)
```

Arguments

df	tidy data_frame retrieved after import of a 2D-TPP dataset, potential filtering and addition of a column "nObs" containing the number of observations per protein
maxit	maximal number of iterations the optimization should be given, default is set to 500
optim_fun	optimization function that should be used for fitting the H0 model
gr_fun	optional gradient function for optim_fun, default is NULL

Value

data frame with H0 model characteristics for each protein

Examples

```
data("simulated_cell_extract_df")
temp_df <- simulated_cell_extract_df %>%
  filter(clustername %in% paste0("protein", 1:5)) %>%
  group_by(representative) %>%
  mutate(nObs = n()) %>%
  ungroup

fitH0Model(temp_df)
```

fitH1Model

Fit H1 model and evaluate fit statistics

Description

Fit H1 model and evaluate fit statistics

Usage

```
fitH1Model(
  df,
  maxit = 500,
  optim_fun = .min_RSS_h1_slope_pEC50,
  optim_fun_2 = NULL,
  gr_fun = NULL,
  gr_fun_2 = NULL,
  ec50_lower_limit = NULL,
  ec50_upper_limit = NULL,
  slopEC50 = TRUE
)
```

Arguments

df	tidy data_frame retrieved after import of a 2D-TPP dataset, potential filtering and addition of a column "nObs" containing the number of observations per protein
maxit	maximal number of iterations the optimization should be given, default is set to 500
optim_fun	optimization function that should be used for fitting the H0 model
optim_fun_2	optional second optimization function for fitting the H1 model that should be used based on the fitted parameters of the optimization for based on optim_fun
gr_fun	optional gradient function for optim_fun, default is NULL
gr_fun_2	optional gradient function for optim_fun_2, default is NULL
ec50_lower_limit	lower limit of ec50 parameter
ec50_upper_limit	lower limit of ec50 parameter
slopEC50	logical flag indicating whether the h1 model is fitted with a linear model describing the shift of the pEC50 over temperatures

Value

data frame with H1 model characteristics for each protein

Examples

```
data("simulated_cell_extract_df")
temp_df <- simulated_cell_extract_df %>%
  filter(clustername %in% paste0("protein", 1:5)) %>%
  group_by(representative) %>%
  mutate(nObs = n()) %>%
  ungroup

fitH1Model(temp_df)
```

getFDR

Get FDR for given F statistics based on true and null dataset

Description

Get FDR for given F statistics based on true and null dataset

Usage

```
getFDR(df_out, df_null, squeezeDenominator = TRUE)
```

Arguments

df_out data frame containing results from analysis by fitAndEvalDataset
df_null data frame containing results from analysis by bootstrapNull
squeezeDenominator logical indicating whether F statistic denominator should be shrunked using limma::squeezeVar

Value

data frame annotating each protein with a FDR based on it's F statistic and number of observations

Examples

```
data("simulated_cell_extract_df")
temp_df <- simulated_cell_extract_df %>%
  filter(clustername %in% paste0("protein", 1:5)) %>%
  group_by(representative) %>%
  mutate(nObs = n()) %>%
  ungroup
example_out <- fitAndEvalDataset(temp_df)
example_null <- bootstrapNull(temp_df, B = 1)
getFDR(example_out, example_null)
```

getModelParamsDf *Get H0 and H1 model parameters*

Description

Get H0 and H1 model parameters

Usage

```
getModelParamsDf(
  df,
  minObs = 20,
  optim_fun_h0 = .min_RSS_h0,
  optim_fun_h1 = .min_RSS_h1_slope_pEC50,
  optim_fun_h1_2 = NULL,
  gr_fun_h0 = NULL,
  gr_fun_h1 = NULL,
  gr_fun_h1_2 = NULL,
  slopEC50 = TRUE,
  maxit = 500,
  qualColName = "qupm"
)
```

Arguments

df	tidy data_frame retrieved after import of a 2D-TPP dataset, potential filtering and addition of a column "nObs" containing the number of observations per protein
minObs	numeric value of minimal number of observations that should be required per protein
optim_fun_h0	optimization function that should be used for fitting the H0 model
optim_fun_h1	optimization function that should be used for fitting the H1 model
optim_fun_h1_2	optional additional optimization function that will be run with parameters retrieved from optim_fun_h1 and should be used for fitting the H1 model with the trimmed sum model, default is NULL
gr_fun_h0	optional gradient function for optim_fun_h0, default is NULL
gr_fun_h1	optional gradient function for optim_fun_h1, default is NULL
gr_fun_h1_2	optional gradient function for optim_fun_h1_2, default is NULL
slopEC50	logical flag indicating whether the h1 model is fitted with a linear model describing the shift of the pEC50 over temperatures
maxit	maximal number of iterations the optimization should be given, default is set to 500
qualColName	name of column indicating quantification quality e.g. number of unique peptides used for quantification, default: "qupm"

Value

a data.frame with fitted null and alternative model parameters

Examples

```
data("simulated_cell_extract_df")
getModelParamsDf(simulated_cell_extract_df)
```

getPEC504Temperature *Get pEC50 for a protein of interest at a specific temperatures (optimally the melting point of the protein)*

Description

Get pEC50 for a protein of interest at a specific temperatures (optimally the melting point of the protein)

Usage

```
getPEC504Temperature(fstat_df, protein, temperaturePEC50 = 60)
```

Arguments

`fstat_df` data frame as obtained after calling `getModelParamsDf`, containing fitted null and alternative model parameters for each protein

`protein` character string referring to the protein of interest

`temperaturePEC50` temperature (numeric) at which pEC50 should be inferred

Value

numeric value specifying the pEC50 for the indicated protein and temperature

Examples

```
data("simulated_cell_extract_df")

model_params_df <- getModelParamsDf(
  df = filter(simulated_cell_extract_df,
             clustertype == "tp1"))

getPEC504Temperature(
  fstat_df = model_params_df,
  protein = "tp1",
  temperaturePEC50 = 60)
```

`getPvalues` *Compute p-values for given F statistics based on true and null dataset*

Description

Compute p-values for given F statistics based on true and null dataset

Usage

```
getPvalues(df_out, df_null, pseudo_count = 1, squeezeDenominator = FALSE)
```

Arguments

`df_out` data frame containing results from analysis by `fitAndEvalDataset`

`df_null` data frame containing results from analysis by `bootstrapNull`

`pseudo_count` numeric larger or equal to 0 added to both counts of protein with an F-statistic higher than a threshold θ of the true and bootstrapped datasets

`squeezeDenominator` logical indicating whether F statistic denominator should be shrunk using `limma::squeezeVar`

Value

data frame annotating each protein with a FDR based on its F statistic and number of observations

Examples

```

data("simulated_cell_extract_df")
temp_df <- simulated_cell_extract_df %>%
  filter(clustername %in% paste0("protein", 1:3)) %>%
  group_by(representative) %>%
  mutate(nObs = n()) %>%
  ungroup
example_out <- fitAndEvalDataset(temp_df)
example_null <- bootstrapNull(temp_df, B = 2)
getPvalues(
  example_out,
  example_null)

```

gg_qq

*Plot qq-plot of true data and bootstrapped null with ggplot***Description**

Plot qq-plot of true data and bootstrapped null with ggplot

Usage

```

gg_qq(
  x,
  y,
  xlab = "F-statistics from sampled Null distr.",
  ylab = "observed F-statistics",
  alpha = 0.25,
  gg_theme = theme_classic(),
  offset = 1,
  plot_diagonal = TRUE
)

```

Arguments

x	vector containing values of values of first distribution to compare
y	vector containing values of values of second distribution to compare
xlab	x-axis label
ylab	y-axis label
alpha	transparency parameter between 0 and 1
gg_theme	ggplot theme, default is theme_classic()
offset	offset for x and y axis on top of maximal values
plot_diagonal	logical parameter indicating whether an identity line should be plotted

Value

A ggplot displaying the qq-plot of a true and a bootstrapped null distribution

Examples

```
data("simulated_cell_extract_df")
recomputeSignalFromRatios(simulated_cell_extract_df)
```

import2dDataset	<i>Import 2D-TPP dataset using a config table</i>
-----------------	---

Description

Import 2D-TPP dataset using a config table

Usage

```
import2dDataset(
  configTable,
  data,
  idVar = "representative",
  intensityStr = "sumionarea_protein_",
  fcStr = "rel_fc_protein_",
  nonZeroCols = "qssm",
  geneNameVar = "clustername",
  addCol = NULL,
  qualColName = "qupm",
  naStrs = c("NA", "n/d", "NaN"),
  concFactor = 1e+06,
  medianNormalizeFC = TRUE,
  filterContaminants = TRUE
)
```

Arguments

configTable	character string of a file path to a config table
data	possible list of datasets from different MS runs corresponding to a 2D-TPP dataset, circumvents loading datasets referencend in config table, default is NULL
idVar	character string indicating which data column provides the unique identifiers for each protein.
intensityStr	character string indicating which columns contain raw intensities measurements
fcStr	character string indicating which columns contain the actual fold change values. Those column names containing the suffix fcStr will be regarded as containing fold change values.

nonZeroCols	column like default qsm that should be imported and requested to be non-zero in analyzed data
geneNameVar	character string of the column name that describes the gene name of a given protein in the raw data files
addCol	character string indicating additional column to import
qualColName	character string indicating which column can be used for additional quality criteria when deciding between different non-unique protein identifiers.
naStrs	character vector indicating missing values in the data table. When reading data from file, this value will be passed on to the argument <code>na.strings</code> in function <code>read.delim</code> .
concFactor	numeric value that indicates how concentrations need to be adjusted to yield total unit e.g. default <code>mmol - 1e6</code>
medianNormalizeFC	perform median normalization (default: TRUE).
filterContaminants	boolean variable indicating whether data should be filtered to exclude contaminants (default: TRUE).

Value

tidy data frame representing a 2D-TPP dataset

Examples

```
data("config_tab")
data("raw_dat_list")
import_df <- import2dDataset(configTable = config_tab,
                             data = raw_dat_list,
                             idVar = "protein_id",
                             intensityStr = "signal_sum_",
                             fcStr = "rel_fc_",
                             nonZeroCols = "qsm",
                             geneNameVar = "gene_name",
                             addCol = NULL,
                             qualColName = "qupm",
                             naStrs = c("NA", "n/d", "NaN"),
                             concFactor = 1e6,
                             medianNormalizeFC = TRUE,
                             filterContaminants = TRUE)
```

import2dMain

Import 2D-TPP dataset main function

Description

Import 2D-TPP dataset main function

Usage

```
import2dMain(
  configTable,
  data,
  idVar,
  fcStr,
  addCol,
  naStrs,
  intensityStr,
  qualColName,
  nonZeroCols
)
```

Arguments

configTable	character string of a file path to a config table
data	possible list of datasets from different MS runs corresponding to a 2D-TPP dataset, circumvents loading datasets referencend in config table, default is NULL
idVar	character string indicating which data column provides the unique identifiers for each protein.
fcStr	character string indicating which columns contain the actual fold change values. Those column names containing the suffix fcStr will be regarded as containing fold change values.
addCol	character string indicating additional column to import
naStrs	character vector indicating missing values in the data table. When reading data from file, this value will be passed on to the argument na.strings in function read.delim.
intensityStr	character string indicating which columns contain raw intensities measurements
qualColName	character string indicating which column can be used for additional quality criteria when deciding between different non-unique protein identifiers.
nonZeroCols	column like default qssm that should be imported and requested to be non-zero in analyzed data

Value

list of data frames containing different datasets

Examples

```
data("config_tab")
data("raw_dat_list")
dataList <- import2dMain(configTable = config_tab,
  data = raw_dat_list,
  idVar = "protein_id",
  fcStr = "rel_fc_",
  addCol = "gene_name",
  naStrs = NA,
```

```
intensityStr = "signal_sum_",
nonZeroCols = "qusm",
qualColName = "qupm")
```

plot2dTppFcHeatmap *Plot heatmap of 2D thermal profile fold changes of a protein of choice*

Description

Plot heatmap of 2D thermal profile fold changes of a protein of choice

Usage

```
plot2dTppFcHeatmap(df, name, drug_name = "", midpoint = 1)
```

Arguments

df	tidy data frame of a 2D-TPP dataset
name	gene name (clustername) of protein that should be visualized
drug_name	character string of profiled drug name
midpoint	midpoint of fold changes for color scaling, default: 1

Value

A ggplot displaying the thermal profile as a heatmap of fold changes of a protein of choice in a dataset of choice

Examples

```
data("simulated_cell_extract_df")
plot2dTppFcHeatmap(simulated_cell_extract_df,
  "tp2", drug_name = "drug1")
```

plot2dTppFit *Plot H0 or H1 fit of 2D thermal profile intensities of a protein of choice*

Description

Plot H0 or H1 fit of 2D thermal profile intensities of a protein of choice

Usage

```
plot2dTppFit(
  df,
  name,
  model_type = "H0",
  optim_fun = .min_RSS_h0,
  optim_fun_2 = NULL,
  maxit = 500,
  xlab = "-log10(conc.)",
  ylab = "log2(summed intensities)",
  dot_size = 1,
  line_type = "solid",
  fit_color = "gray30"
)
```

Arguments

df	tidy data frame of a 2D-TPP dataset
name	gene name (clustername) of protein that should be visualized
model_type	character string indicating whether the "H0" or the "H1" model should be fitted
optim_fun	optimization function that should be used for fitting either the H0 or H1 model
optim_fun_2	optional additional optimization function that will be run with paramters retrieved from optim_fun and should be used for fitting the H1 model with the trimmed sum model, default is NULL
maxit	maximal number of iterations the optimization should be given, default is set to 500
xlab	character string of x-axis label of plot
ylab	character string of y-axis label of plot
dot_size	numeric indicating the size of the data points to plot
line_type	character string defining the line type of the fitted curve, default "dashed"
fit_color	character string defining the color of the fitted curve

Value

A ggplot displaying the thermal profile of a protein of choice in a dataset of choice

Examples

```
data("simulated_cell_extract_df")
plot2dTppProfile(simulated_cell_extract_df, "protein1")
```

plot2dTppProfile	<i>Plot 2D thermal profile intensities of a protein of choice</i>
------------------	---

Description

Plot 2D thermal profile intensities of a protein of choice

Usage

```
plot2dTppProfile(df, name)
```

Arguments

df	tidy data frame of a 2D-TPP dataset
name	gene name (clustername) of protein that should be visualized

Value

A ggplot displaying the thermal profile of a protein of choice in a dataset of choice

Examples

```
data("simulated_cell_extract_df")  
plot2dTppProfile(simulated_cell_extract_df, "protein1")
```

plot2dTppRelProfile	<i>Plot 2D thermal profile ratios of a protein of choice</i>
---------------------	--

Description

Plot 2D thermal profile ratios of a protein of choice

Usage

```
plot2dTppRelProfile(df, name)
```

Arguments

df	tidy data frame of a 2D-TPP dataset
name	gene name (clustername) of protein that should be visualized

Value

A ggplot displaying the thermal profile ratios of a protein of choice in a dataset of choice

Examples

```
data("simulated_cell_extract_df")
plot2dTppRelProfile(simulated_cell_extract_df, "protein1")
```

plot2dTppVolcano *Plot Volcano plot of TPP2D results*

Description

Plot Volcano plot of TPP2D results

Usage

```
plot2dTppVolcano(
  fdr_df,
  hits_df,
  alpha = 0.5,
  title_string = "",
  x_lim = NULL,
  y_lim = NULL,
  facet_by_obs = FALSE
)
```

Arguments

fdr_df	data frame obtained from 'getFDR'
hits_df	hits_df data frame obtained from 'findHits'
alpha	transparency level of plotted points
title_string	character argument handed over to ggtitle
x_lim	vector with two numerics indicating the x axis limits
y_lim	vector with two numerics indicating the y axis limits
facet_by_obs	logical indicating whether plot should be faceted by number of observations, default: FALSE

Value

a ggplot displaying a volcano plot of the results obtained after a TPP2D analysis

Examples

```

data("simulated_cell_extract_df")
temp_df <- simulated_cell_extract_df %>%
  filter(clustername %in% paste0("protein", 1:5)) %>%
  group_by(representative) %>%
  mutate(nObs = n()) %>%
  ungroup
example_params <- getModelParamsDf(temp_df)
example_fstat <- computeFStatFromParams(example_params)
example_null <- bootstrapNullAlternativeModel(
  df = temp_df, params_df = example_params,
  B = 2)
fdr_df <- getFDR(example_fstat, example_null)
hits_df <- findHits(fdr_df, 0.1)
plot2dTppVolcano(fdr_df = fdr_df, hits_df = hits_df)

```

raw_dat_list	<i>Example raw data for a subset of a simulated 2D-TPP cell extract dataset</i>
--------------	---

Description

Simulated example dataset obtained by 2D-TPP experiments for analysis by the TPP2D-package. It contains a list of data frames resembling raw data files returned from a MS database search with 200 simulated protein profiles (protein1-200) and 3 spiked-in true positives (TP1-3).

Usage

```
data("raw_dat_list")
```

Format

list of data frames with columns representative (protein id), clustername (gene name), temperature, log_conc, raw_value, rel_value, value and log2_value

recomputeSignalFromRatios	<i>Recompute robust signal intensities based on bootstrapped TMT channel ratios</i>
---------------------------	---

Description

Recompute robust signal intensities based on bootstrapped TMT channel ratios

Usage

```
recomputeSignalFromRatios(df)
```

Arguments

df tidy data_frame retrieved after import of a 2D-TPP dataset

Value

A data_frame with recomputed signal intensities (columnname: value) and log2 transformed signal intensities (columnname: log2_value) that more reliably reflect relative ratios between the TMT channels

Examples

```
data("simulated_cell_extract_df")
recomputeSignalFromRatios(simulated_cell_extract_df)
```

renameColumns	<i>Rename columns of imported data frame</i>
---------------	--

Description

Rename columns of imported data frame

Usage

```
renameColumns(dataLong, idVar, geneNameVar)
```

Arguments

dataLong long format data frame of imported dataset

idVar character string indicating which data column provides the unique identifiers for each protein.

geneNameVar character string of the column name that describes the gene name of a given protein in the raw data files

Value

data frame containing imported data with renamed columns

Examples

```
data("config_tab")
data("raw_dat_list")

dataList <- import2dMain(configTable = config_tab,
                        data = raw_dat_list,
                        idVar = "protein_id",
                        fcStr = "rel_fc_",
                        addCol = "gene_name",
```

```
      naStrs = NA,
      intensityStr = "signal_sum_",
      nonZeroCols = "qusm",
      qualColName = "qupm")
configLong <- configWide2Long(configWide = config_tab)
annoDat <- annotateDataList(dataList = dataList,
                           geneNameVar = "gene_name",
                           configLong = configLong,
                           intensityStr = "signal_sum_",
                           fcStr = "rel_fc_")
renameColumns(annoDat,
              idVar = "protein_id",
              geneNameVar = "gene_name")
```

resolveAmbiguousProteinNames

Resolve ambiguous protein names

Description

Resolve ambiguous protein names

Usage

```
resolveAmbiguousProteinNames(df, includeIsoforms = FALSE)
```

Arguments

df tidy data_frame retrieved after import of a 2D-TPP dataset

includeIsoforms logical indicating whether protein isoform should be kept for analysis

Value

data frame with resolved protein name ambiguity

Examples

```
tst_df <- bind_rows(tibble(representative = rep(1:3, each = 3),
                          clustername = rep(letters[1:3], each = 3)),
                  tibble(representative = rep(c(4, 5), c(3, 2)),
                          clustername = rep(c("a", "b"), c(3, 2))))

resolveAmbiguousProteinNames(tst_df)
```

runTPP2D	<i>Run complete TPP2D analysis</i>
----------	------------------------------------

Description

Run complete TPP2D analysis

Usage

```
runTPP2D(  
  df = NULL,  
  configTable = NULL,  
  data = NULL,  
  idVar = "protein_id",  
  intensityStr = "signal_sum",  
  fcStr = "rel_fc",  
  nonZeroCols = "qusm",  
  geneNameVar = "gene_name",  
  addCol = NULL,  
  qualColName = "qupm",  
  naStrs = c("NA", "n/d", "NaN"),  
  concFactor = 1e+06,  
  medianNormalizeFC = TRUE,  
  filterContaminants = TRUE,  
  recomputeSignalRatios = FALSE,  
  minObs = 20,  
  independentFiltering = FALSE,  
  fcThres = 1.5,  
  optim_fun_h0 = .min_RSS_h0,  
  optim_fun_h1 = .min_RSS_h1_slope_pEC50,  
  optim_fun_h1_2 = NULL,  
  gr_fun_h0 = NULL,  
  gr_fun_h1 = NULL,  
  gr_fun_h1_2 = NULL,  
  slopEC50 = TRUE,  
  maxit = 750,  
  BPPARAM = BiocParallel::SerialParam(progressbar = TRUE),  
  B = 20,  
  byMsExp = TRUE,  
  alpha = 0.1  
)
```

Arguments

df	tidy data_frame retrieved after import of a 2D-TPP dataset, potential filtering and addition of a column "nObs" containing the number of observations per protein
----	---

configTable	character string of a file path to a config table
data	possible list of datasets from different MS runs corresponding to a 2D-TPP dataset, circumvents loading datasets referencend in config table, default is NULL
idVar	character string indicating which data column provides the unique identifiers for each protein.
intensityStr	character string indicating which columns contain raw intensities measurements
fcStr	character string indicating which columns contain the actual fold change values. Those column names containing the suffix fcStr will be regarded as containing fold change values.
nonZeroCols	column like default qssm that should be imported and requested to be non-zero in analyzed data
geneNameVar	character string of the column name that describes the gene name of a given protein in the raw data files
addCol	character string indicating additional column to import
qualColName	character string indicating which column can be used for additional quality criteria when deciding between different non-unique protein identifiers.
naStrs	character vector indicating missing values in the data table. When reading data from file, this value will be passed on to the argument na.strings in function read.delim.
concFactor	numeric value that indicates how concentrations need to be adjusted to yield total unit e.g. default mmol - 1e6
medianNormalizeFC	perform median normalization (default: TRUE).
filterContaminants	logical variable indicating whether data should be filtered to exclude contaminants (default: TRUE).
recomputeSignalRatios	logical variable indicaitng whether signals should be recomputed from relative fold changes, recommended if Isobarquant was used for protein quantification
minObs	number of minimal observations per protein to include it in the analysis
independentFiltering	logical variable indicating whether independent filtering should be performed based on minimal fold changes per protein profile
fcThres	numeric value of minimal fold change (or inverse fold change) a protein has to show to be kept upon independent filtering
optim_fun_h0	optimization function that should be used for fitting the H0 model
optim_fun_h1	optimization function that should be used for fitting the H1 model
optim_fun_h1_2	optional additional optimization function that will be run with paramters retrieved from optim_fun_h1 and should be used for fitting the H1 model with the trimmed sum model, default is NULL
gr_fun_h0	optional gradient function for optim_fun_h0, default is NULL
gr_fun_h1	optional gradient function for optim_fun_h1, default is NULL

gr_fun_h1_2	optional gradient function for optim_fun_h1_2, default is NULL
slopEC50	logical flag indicating whether the h1 model is fitted with a linear model describing the shift of the pEC50 over temperatures
maxit	maximal number of iterations the optimization should be given, default is set to 500
BPPARAM	= BiocParallel::SerialParam(progressbar = TRUE),
B	numeric value indicating number of rounds of bootstraps that should be performed to estimate the null distribution
byMsExp	logical indicating whether bootstrapping should be performed within MS experiments
alpha	FDR level that should be controlled

Value

a tpp2dExperiment object

Examples

```
data("simulated_cell_extract_df")
runTPP2D(df = simulated_cell_extract_df %>%
  filter(representative %in% 1:3),
  B = 1)
```

simulated_cell_extract_df

Example subset of a simulated 2D-TPP cell extract dataset

Description

Simulated example dataset obtained by 2D-TPP experiments for analysis by the TPP2D-package. It contains a tidy data frame after import and recomputing of robust signal intensities with 200 simulated protein profiles (protein1-200) and 3 spiked-in true positives (TP1-3)

Usage

```
data("simulated_cell_extract_df")
```

Format

data frame with columns representative (protein id), clustername (gene name), temperature, log_conc, raw_value, rel_value, value and log2_value

TPP2D-defunct

*Defunct functions in package **TPP2D**.*

Description

The functions listed below are defunct and will be removed in the near future. When possible, alternative functions with similar functionality are also mentioned. Help pages for deprecated functions are available at `help("-deprecated")`.

Usage

```
computeFdr(df_out, df_null)
```

Details

TPP2D defunct functions

`computeFdr`

For `computeFdr`, use [getFDR](#).

`tpp2dExperiment-class` *S4 TPP2D Experiment Class*

Description

S4 TPP2D Experiment Class

Value

an object of class `tpp2dExperiment`

Slots

`configTable` data.frame.

`idVar` character.

`intensityStr` character.

`fcStr` character.

`nonZeroCols` character.

`geneNameVar` character.

`qualColName` character.

`naStrs` character.

`concFactor` numeric.

medianNormalizeFC logical.
 filterContaminants logical.
 minObs numeric.
 independentFiltering logical.
 fcThres numeric.
 optim_fun_h0 function.
 optim_fun_h1 function.
 slopEC50 logical.
 maxit numeric.
 BPPARAM character.
 B numeric
 byMsExp logical.
 alpha numeric.
 tidyDataTable data.frame.
 modelParamsDf data.frame
 resultTable data.frame
 bootstrapNullDf data.frame
 hitTable data.frame

Examples

```
tpp2dObj <- new("tpp2dExperiment")
```

TPP_importCheckConfigTable

Import and check configuration table

Description

Import and check configuration table

Usage

```
TPP_importCheckConfigTable(infoTable, type = "2D")
```

Arguments

infoTable	character string of a file path to a config table (excel,txt or csv file) or data frame containing a config table
type	character string indicating dataset type default is 2D

Value

data frame with config table

Examples

```
data("config_tab")
TPP_importCheckConfigTable(config_tab, type = "2D")
```

Index

- * **datasets**
 - [config_tab, 12](#)
 - [raw_dat_list, 29](#)
 - [simulated_cell_extract_df, 34](#)
- * **internal**
 - [computeFdr-defunct, 10](#)
 - [TPP2D-defunct, 35](#)
- [annotateDataList, 3](#)
- [bootstrapNull, 4](#)
- [bootstrapNullAlternativeModel, 5](#)
- [bootstrapNullAlternativeModelFast, 7](#)
- [competeModels, 9](#)
- [computeFdr \(TPP2D-defunct\), 35](#)
- [computeFdr-defunct, 10](#)
- [computeFstat, 10](#)
- [computeFStatFromParams, 11](#)
- [config_tab, 12](#)
- [configWide2Long, 12](#)
- [filterOutContaminants, 13](#)
- [findHits, 13](#)
- [fitAndEvalDataset, 14](#)
- [fitH0Model, 15](#)
- [fitH1Model, 16](#)
- [getFDR, 17, 35](#)
- [getModelParamsDf, 18](#)
- [getPEC504Temperature, 19](#)
- [getPvalues, 20](#)
- [gg_qq, 21](#)
- [import2dDataset, 22](#)
- [import2dMain, 23](#)
- [plot2dTppFcHeatmap, 25](#)
- [plot2dTppFit, 25](#)
- [plot2dTppProfile, 27](#)
- [plot2dTppRelProfile, 27](#)
- [plot2dTppVolcano, 28](#)
- [raw_dat_list, 29](#)
- [recomputeSignalFromRatios, 29](#)
- [renameColumns, 30](#)
- [resolveAmbiguousProteinNames, 31](#)
- [runTPP2D, 32](#)
- [simulated_cell_extract_df, 34](#)
- [tpcaResult \(tpp2dExperiment-class\), 35](#)
- [TPP2D-defunct, 35](#)
- [tpp2dExperiment-class, 35](#)
- [TPP_importCheckConfigTable, 36](#)