

# Package ‘RNAmodR’

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**Type** Package

**Title** Detection of post-transcriptional modifications in high throughput sequencing data

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**Description** RNAmodR provides classes and workflows for loading/aggregation data from high throughput sequencing aimed at detecting post-transcriptional modifications through analysis of specific patterns. In addition, utilities are provided to validate and visualize the results. The RNAmodR package provides a core functionality from which specific analysis strategies can be easily implemented as a separate package.

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'Modifier-utils.R' 'SequenceDataFrame-class.R'  
'normalization.R' 'SequenceData-class.R'  
'SequenceDataSet-class.R' 'SequenceDataList-class.R'  
'Modifier-class.R' 'ModifierSet-class.R'  
'Modifier-Inosine-class.R' 'Modifier-Inosine-viz.R'  
'Modifier-roc.R' 'Modifier-subset.R' 'Modifier-viz.R'  
'ModifierSet-comparison.R' 'ModifierSet-viz.R'  
'RNAmodR-external-functions.R' 'RNAmodR-summary.R'  
'SequenceData-coverage.R' 'SequenceData-end-pos.R'

'SequenceData-normalized-end-pos.R' 'SequenceData-pileup.R'  
 'SequenceData-viz.R' 'SequenceData-protected-end-pos.R'  
 'SequenceData-stats.R' 'SequenceData-subset.R'  
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 'utils.R'

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RNAmodR-package	<i>RNAmodR: Detection of post-transcriptional modifications in high throughput sequencing data</i>
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**Description**

RNAmodR provides classes and workflows for loading/aggregation data from high throughput sequencing aimed at detecting post-transcriptional modifications through analysis of specific patterns. In addition, utilities are provided to validate and visualize the results. The RNAmodR package provides a core functionality from which specific analysis strategies can be easily implemented as a separate package.

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**See Also**

Useful links:

- <https://github.com/FelixErnst/RNAmodR>
- Report bugs at <https://github.com/FelixErnst/RNAmodR/issues>

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aggregate	<i>Aggregate data per positions</i>
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**Description**

The aggregate function is defined for each [SequenceData](#) object and can be used directly on a [SequenceData](#) object or indirectly via a [Modifier](#) object.

For the letter the call is redirect to the [SequenceData](#) object, the result summarized as defined for the individual [Modifier](#) class and stored in the aggregate slot of the [Modifier](#) object. The data is then used for subsequent tasks, such as search for modifications and visualization of the results.

The summarization is implemented in the aggregateData for each type of [Modifier](#) class. The stored data from the aggregate slot can be retrieved using the getAggregateData function.

Whether the aggregated data is already present in the aggregate slot can be checked using the hasAggregateData function.

For SequenceDataSet, SequenceDataList and ModifierSet classes wrapper of the aggregate function exist as well.

**Usage**

```

aggregate(x, ...)

aggregateData(x, ...)

getAggregateData(x)

hasAggregateData(x)

## S4 method for signature 'SequenceData'
aggregate(x, condition = c())

## S4 method for signature 'SequenceData'
aggregateData(x, condition)

## S4 method for signature 'SequenceDataSet'
aggregate(x, condition = "Treated")

## S4 method for signature 'SequenceDataList'
aggregate(x, condition = "Treated")

## S4 method for signature 'Modifier'
aggregate(x, force = FALSE)

## S4 method for signature 'Modifier'
aggregateData(x)

## S4 method for signature 'Modifier'
getAggregateData(x)

## S4 method for signature 'Modifier'
hasAggregateData(x)

## S4 method for signature 'ModifierSet'
aggregate(x, force = FALSE)

```

**Arguments**

x	a <a href="#">SequenceData</a> , <a href="#">SequenceDataSet</a> , <a href="#">SequenceDataList</a> , <a href="#">Modifier</a> or <a href="#">ModifierSet</a> object.
...	additional arguments
condition	character value, which selects, for which condition the data should be aggregated. One of the following values: Both, Control, Treated
force	whether to recreate the aggregated data, if it is already stored inside the <a href="#">Modifier</a> object.

**Value**

- `aggregate`: for [SequenceData](#) object the aggregated data is returned as a [SplitDataFrameList](#) with an element per transcript, whereas for a [Modifier](#) the modified input object is returned, containing the aggregated data, which can be accessed using `getAggregateData`.

- `getAggregateData`: only for `Modifier`: a `SplitDataFrameList` with an element per transcript is returned. If the aggregated data is not stored in the object, it is generated on the fly, but does not persist.
- `hasAggregateData`: `TRUE` or `FALSE`. Does the `Modifier` object already contain aggregated data?

If 'x' is a

- `SequenceData` a `SplitDataFrameList` with elements per transcript.
- `SequenceDataSet` or `SequenceDataList` a `SimpleList` with `SplitDataFrameList` as elements.
- `Modifier` or `ModifierSet` an updated `Modifier` object. The data can be accessed by using the `aggregateData` function.

### Examples

```
data(e5sd,package="RNAmodR")
data(msi,package="RNAmodR")
# modify() triggers the search for modifications in the data contained in
# the Modifier or ModifierSet object
sdf1 <- aggregate(e5sd)
mi <- aggregate(msi[[1]])
```

---

compare

*Comparison of Samples*

---

### Description

To compare data of different samples, a `ModifierSet` can be used. To select the data alongside the transcripts and their positions a `GRanges` or a `GRangesList` needs to be provided. In case of a `GRanges` object, the parent column must match the transcript names as defined by the output of `ranges(x)`, whereas in case of a `GRangesList` the element names must match the transcript names.

### Usage

```
compare(x, name, pos = 1L, ...)

compareByCoord(x, coord, ...)

plotCompare(x, name, pos = 1L, normalize, ...)

plotCompareByCoord(x, coord, normalize, ...)

## S4 method for signature 'ModifierSet'
compare(x, name, pos = 1L, normalize, ...)

## S4 method for signature 'ModifierSet,GRanges'
compareByCoord(x, coord, normalize, ...)

## S4 method for signature 'ModifierSet,GRangesList'
compareByCoord(x, coord, normalize, ...)
```

```
## S4 method for signature 'ModifierSet'
plotCompare(x, name, pos = 1L, normalize, ...)

## S4 method for signature 'ModifierSet,GRanges'
plotCompareByCoord(x, coord, normalize, ...)

## S4 method for signature 'ModifierSet,GRangesList'
plotCompareByCoord(x, coord, normalize, ...)
```

## Arguments

x	a Modifier or ModifierSet object.
name	Only for compare: the transcript name
pos	Only for compare: pos for comparison
...	optional parameters: <ul style="list-style-type: none"> <li>• alias a data.frame with two columns, tx_id and name, to convert transcript ids to another identifier</li> <li>• name Limit results to one specific gene or transcript</li> <li>• sequenceData TRUE or FALSE? Should the aggregate of sequenceData be used for the comparison instead of the aggregate data if each Modifier element? (default: sequenceData = FALSE)</li> <li>• compareType a valid score type to use for the comparison. If sequenceData = FALSE this defaults to mainScore(x), whereas if sequenceData = TRUE all columns will be used by setting allTypes = TRUE.</li> <li>• allTypes TRUE or FALSE? Should all available score be compared? (default: allTypes = sequenceData)</li> <li>• ... passed on to <a href="#">subsetByCoord</a></li> </ul>
coord	coordinates of position to subset to. Either a GRanges or a GRangesList object. For both types the 'Parent' column is expected to match the transcript name. The GRangesList object is unlisted and only non duplicated entries are retained.
normalize	either a single logical or character value. If it is a character, it must match one of the names in the ModifierSet.

## Value

compareByCoord returns a [DataFrame](#) and plotCompareByCoord returns a ggplot object, which can be modified further. The DataFrame contains columns per sample as well as the columns names, positions and mod incorporated from the coord input. If coord contains a column Activity this is included in the results as well.

## Examples

```
data(msi, package="RNAmodR")
# constructing a GRanges object to mark positive positions
mod <- modifications(msi)
coord <- unique(unlist(mod))
coord$score <- NULL
coord$sd <- NULL
# return a DataFrame
compareByCoord(msi, coord)
# plot the comparison as a heatmap
plotCompareByCoord(msi, coord)
```

---

CoverageSequenceData-class  
*CoverageSequenceData*

---

## Description

CoverageSequenceData implements [SequenceData](#) to contain and aggregate the coverage of reads per position along the transcripts.

CoverageSequenceData contains one column per data file named using the following naming convention `coverage.condition.replicate`.

`aggregate` calculates the mean and sd for samples in the control and treated condition separately.

## Usage

```
CoverageSequenceDataFrame(
  df,
  ranges,
  sequence,
  replicate,
  condition,
  bamfiles,
  seqinfo
)

CoverageSequenceData(bamfiles, annotation, sequences, seqinfo, ...)

## S4 method for signature
## 'CoverageSequenceData,BamFileList,GRangesList,XStringSet,ScanBamParam'
getData(x, bamfiles, grl, sequences, param, args)

## S4 method for signature 'CoverageSequenceData'
aggregateData(x, condition = c("Both", "Treated", "Control"))

## S4 method for signature 'CoverageSequenceData'
getDataTrack(x, name, ...)
```

## Arguments

`df, ranges, sequence, replicate`  
 inputs for creating a `SequenceDataFrame`. See [SequenceDataFrame](#).

`condition` For `aggregate`: condition for which the data should be aggregated.

`bamfiles, annotation, seqinfo, grl, sequences, param, args, ...`  
 See [SequenceData](#)

`x` a `CoverageSequenceData`

`name` For `getDataTrack`: a valid transcript name. Must be a name of `ranges(x)`

## Value

a `CoverageSequenceData` object

**Examples**

```
# Construction of a CoverageSequenceData objectobject
library(RNAmoR.Data)
library(rtracklayer)
annotation <- GFF3File(RNAmoR.Data.example.man.gff3())
sequences <- RNAmoR.Data.example.man.fasta()
files <- c(treated = RNAmoR.Data.example.wt.1())
csd <- CoverageSequenceData(files, annotation = annotation,
                           sequences = sequences)
```

---

EndSequenceData-class *End5SequenceData/End3SequenceData/EndSequenceData*

---

**Description**

The End5SequenceData/End3SequenceData/EndSequenceData classes aggregate the counts of read ends at each position along a transcript. End5SequenceData/End3SequenceData classes aggregate either the 5'-end or 3'-end, the EndSequenceData aggregates both.

All three classes contain one column per data file named using the following naming convention (end5/end3/end).condition.replicate.

aggregate calculates the mean and sd for samples in the control and treated condition separately.

**Usage**

```
End5SequenceDataFrame(
  df,
  ranges,
  sequence,
  replicate,
  condition,
  bamfiles,
  seqinfo
)
```

```
End3SequenceDataFrame(
  df,
  ranges,
  sequence,
  replicate,
  condition,
  bamfiles,
  seqinfo
)
```

```
EndSequenceDataFrame(
  df,
  ranges,
  sequence,
  replicate,
  condition,
```

```

    bamfiles,
    seqinfo
)

End5SequenceData(bamfiles, annotation, sequences, seqinfo, ...)

End3SequenceData(bamfiles, annotation, sequences, seqinfo, ...)

EndSequenceData(bamfiles, annotation, sequences, seqinfo, ...)

## S4 method for signature
## 'End5SequenceData,BamFileList,GRangesList,XStringSet,ScanBamParam'
getData(x, bamfiles, grl, sequences, param, args)

## S4 method for signature
## 'End3SequenceData,BamFileList,GRangesList,XStringSet,ScanBamParam'
getData(x, bamfiles, grl, sequences, param, args)

## S4 method for signature
## 'EndSequenceData,BamFileList,GRangesList,XStringSet,ScanBamParam'
getData(x, bamfiles, grl, sequences, param, args)

## S4 method for signature 'End5SequenceData'
aggregateData(x, condition = c("Both", "Treated", "Control"))

## S4 method for signature 'End3SequenceData'
aggregateData(x, condition = c("Both", "Treated", "Control"))

## S4 method for signature 'EndSequenceData'
aggregateData(x, condition = c("Both", "Treated", "Control"))

## S4 method for signature 'EndSequenceData'
getDataTrack(x, name, ...)

## S4 method for signature 'End5SequenceData'
getDataTrack(x, name, ...)

## S4 method for signature 'End3SequenceData'
getDataTrack(x, name, ...)

```

## Arguments

df, ranges, sequence, replicate  
 inputs for creating a SequenceDataFrame. See [SequenceDataFrame](#).

condition      For [aggregate](#): condition for which the data should be aggregated.

bamfiles, annotation, seqinfo, grl, sequences, param, args, ...  
 See [SequenceData](#) and [SequenceData-functions](#)

x                a End5SequenceData, End3SequenceData or EndSequenceData object

name            For [getDataTrack](#): a valid transcript name. Must be a name of ranges(x).

**Value**

a End5SequenceData, a End3SequenceData or a EndSequenceData object

**Examples**

```
# Construction of a End5SequenceData object
library(RNAmoR.Data)
library(rtracklayer)
annotation <- GFF3File(RNAmoR.Data.example.man.gff3())
sequences <- RNAmoR.Data.example.man.fasta()
files <- c(treated = RNAmoR.Data.example.wt.1())
e5sd <- End5SequenceData(files, annotation = annotation,
                        sequences = sequences)
```

---

Modifier-class

*The Modifier class*

---

**Description**

The Modifier class is a virtual class, which provides the central functionality to search for post-transcriptional RNA modification patterns in high throughput sequencing data.

Each subclass has to implement the following functions:

- Slot nucleotide: Either "RNA" or "DNA". For convenience the subclasses RNAModifier and DNAModifier are already available and can be inherited from.
- Function `aggregateData`: used for specific data aggregation
- Function `findMod`: used for specific search for modifications

Optionally the function `settings<-` can be implemented to store additional arguments, which the base class does not recognize.

Modifier objects are constructed centrally by calling `Modifier()` with a `className` matching the specific class to be constructed. This will trigger the immediate analysis, if `find.mod` is not set to FALSE.

**Usage**

```
Modifier(className, x, annotation, sequences, seqinfo, ...)
```

```
## S4 method for signature 'SequenceData'
```

```
Modifier(
  className,
  x,
  annotation = NULL,
  sequences = NULL,
  seqinfo = NULL,
  ...
)
```

```
## S4 method for signature 'SequenceDataSet'
```

```
Modifier(
```

```
    className,  
    x,  
    annotation = NULL,  
    sequences = NULL,  
    seqinfo = NULL,  
    ...  
  )  
  
## S4 method for signature 'SequenceDataList'  
Modifier(  
  className,  
  x,  
  annotation = NULL,  
  sequences = NULL,  
  seqinfo = NULL,  
  ...  
)  
  
## S4 method for signature 'character'  
Modifier(  
  className,  
  x,  
  annotation = NULL,  
  sequences = NULL,  
  seqinfo = NULL,  
  ...  
)  
  
## S4 method for signature 'list'  
Modifier(  
  className,  
  x,  
  annotation = NULL,  
  sequences = NULL,  
  seqinfo = NULL,  
  ...  
)  
  
## S4 method for signature 'BamFileList'  
Modifier(  
  className,  
  x,  
  annotation = NULL,  
  sequences = NULL,  
  seqinfo = NULL,  
  ...  
)
```

### Arguments

className	The name of the class which should be constructed.
x	the input which can be of the following types

	<ul style="list-style-type: none"> <li>• <code>SequenceData</code>: a single <code>SequenceData</code> or a list containing only <code>SequenceData</code> objects. The input will just be used to file the data slot of the <code>Modifier</code> and must match the requirements of specific <code>Modifier</code> class.</li> <li>• <code>BamFileList</code>: a named <code>BamFileList</code></li> <li>• <code>character</code>: a character vector, which must be coercible to a named <code>BamFileList</code> referencing existing bam files. Valid names are <code>control</code> and <code>treated</code> to define conditions and replicates</li> </ul>
<code>annotation</code>	annotation data, which must match the information contained in the BAM files. This parameter is only required if <code>x</code> is not a <code>SequenceData</code> object or a list of <code>SequenceData</code> objects.
<code>sequences</code>	sequences matching the target sequences the reads were mapped onto. This must match the information contained in the BAM files. This parameter is only required if <code>x</code> is not a <code>SequenceData</code> object or a list of <code>SequenceData</code> objects.
<code>seqinfo</code>	An optional <code>Seqinfo</code> argument or character vector, which can be coerced to one, to subset the sequences to be analyzed on a per chromosome basis.
<code>...</code>	<p>Additional optional parameters:</p> <ul style="list-style-type: none"> <li>• <code>find.mod</code>: TRUE or FALSE: should the search for modifications be triggered upon construction? If not the search can be started by calling the <code>modify()</code> function.</li> <li>• additional parameters depending on the specific <code>Modifier</code> class</li> </ul> <p>All additional options must be named and will be passed to the <code>settings</code> function and onto the <code>SequenceData</code> objects, if <code>x</code> is not a <code>SequenceData</code> object or a list of <code>SequenceData</code> objects.</p>

### Value

a `Modifier` object of type `className`

### Slots

`nucleotide` a character value, which needs to contain "RNA" or "DNA"

`mod` a character value, which needs to contain one or more elements from the alphabet of a `ModRNAString` or `ModDNAString` class.

`score` the main score identifier used for visualizations

`dataType` the class name(s) of the `SequenceData` class used

`bamfiles` the input bam files as `BamFileList`

`condition` conditions along the `BamFileList`: Either `control` or `treated`

`replicate` replicate number along the `BamFileList` for each of the condition types.

`data` The sequence data object: Either a `SequenceData`, `SequenceDataSet` or a `SequenceDataList` object, if more than one `dataType` is used.

`aggregate` the aggregated data as a `SplitDataFrameList`

`modifications` the found modifications as a `GRanges` object

`settings` arguments used for the analysis as a list

`aggregateValidForCurrentArguments` TRUE or FALSE whether the aggregate data was constructed with the current arguments

`modificationsValidForCurrentArguments` TRUE or FALSE whether the modifications were found with the current arguments

## Creation

Modifier objects can be created in two ways, either by providing a list of bamfiles or SequenceData/SequenceDataSet/SequenceDataList objects, which match the structure in dataType().

dataType() can be a character vector or a list of character vectors and depending on this the input files have to follow this structure:

- a single character: a SequenceData is constructed/expected.
- a character vector: a SequenceDataSet is constructed/expected.
- a list of character vectors: a SequenceDataList is constructed/expected.

The cases for a SequenceData or SequenceDataSet are straight forward, since the input remains the same. The last case is special, since it is a hypothetical option, in which bam files from two or more different methods have to be combined to reliably detect a single modification (The elements of a SequenceDataList don't have to be created from the bamfiles, whereas from a SequenceDataSet they have to be).

For this example a list of character vectors is expected. Each element must be named according to the names of dataType() and contain a character vector for creating a SequenceData object.

All additional options must be named and will be passed to the `settings` function and onto the SequenceData objects, if x is not a SequenceData object or a list of SequenceData objects.

---

Modifier-functions      *Modifier/ModifierSet functions*

---

## Description

For the Modifier and ModifierSet classes a number of functions are implemented to access the data stored by the object.

The validAggregate and validModification functions check if `settings` have been modified, after the data was loaded. This potentially invalidates them. To update the data, run the aggregate or the modify function.

## Usage

bamfiles(x)

mainScore(x)

modifierType(x)

modType(x)

dataType(x)

sequenceData(x)

sequences(x, ...)

validAggregate(x)

```
validModification(x)

## S4 method for signature 'Modifier'
show(object)

## S4 method for signature 'Modifier'
bamfiles(x)

## S4 method for signature 'Modifier'
conditions(object)

## S4 method for signature 'Modifier'
mainScore(x)

## S4 method for signature 'Modifier'
modifierType(x)

## S4 method for signature 'Modifier'
modType(x)

## S4 method for signature 'Modifier'
dataType(x)

## S4 method for signature 'Modifier'
names(x)

## S4 method for signature 'Modifier'
ranges(x)

## S4 method for signature 'Modifier'
replicates(x)

## S4 method for signature 'Modifier'
seqinfo(x)

## S4 method for signature 'Modifier'
seqtype(x)

## S4 method for signature 'Modifier'
sequenceData(x)

## S4 method for signature 'Modifier'
sequences(x, modified = FALSE)

## S4 method for signature 'Modifier'
validAggregate(x)

## S4 method for signature 'Modifier'
validModification(x)

## S4 method for signature 'ModifierSet'
show(object)
```

```

## S4 method for signature 'ModifierSet'
bamfiles(x)

## S4 method for signature 'ModifierSet'
conditions(object)

## S4 method for signature 'ModifierSet'
mainScore(x)

## S4 method for signature 'ModifierSet'
modifications(x, perTranscript = FALSE)

## S4 method for signature 'ModifierSet'
modifierType(x)

## S4 method for signature 'ModifierSet'
modType(x)

## S4 method for signature 'ModifierSet'
dataType(x)

## S4 method for signature 'ModifierSet'
ranges(x)

## S4 method for signature 'ModifierSet'
replicates(x)

## S4 method for signature 'ModifierSet'
seqinfo(x)

## S4 method for signature 'ModifierSet'
seqtype(x)

## S4 method for signature 'ModifierSet'
sequences(x, modified = FALSE)

```

### Arguments

<code>x, object</code>	a <code>Modifier</code> or <code>ModifierSet</code> class
<code>...</code>	Additional arguments.
<code>modified</code>	For sequences: TRUE or FALSE: Should the sequences be returned as a <code>ModRNAString/ModDNAString</code> with the found modifications added on top of the <code>RNAString/ DNAString</code> ? See <a href="#">combineIntoModstrings</a> .
<code>perTranscript</code>	TRUE or FALSE: Should the positions shown per transcript? (default: <code>perTranscript = FALSE</code> )

### Value

- `modifierType`: a character vector with the appropriate class Name of a `Modifier`.
- `modType`: a character vector with the modifications detected by the `Modifier` class.

- seqtype: a single character value defining if either "RNA" or "DNA" modifications are detected by the Modifier class.
- mainScore: a character vector.
- sequenceData: a SequenceData object.
- modifications: a GRanges or GRangesList object describing the found modifications.
- seqinfo: a Seqinfo object.
- sequences: a RNASTingSet object.
- ranges: a GRangesList object with each element per transcript.
- bamfiles: a BamFileList object.
- validAggregate: TRUE or FALSE. Checks if current settings are the same for which the data was aggregate
- validModification: TRUE or FALSE. Checks if current settings are the same for which modification were found

### See Also

[settings](#)

### Examples

```
data(msi,package="RNAmodR")
mi <- msi[[1]]
modifierType(mi) # The class name of the Modifier object
modifierType(msi)
seqtype(mi)
modType(mi)
mainScore(mi)
sequenceData(mi)
modifications(mi)
# general accessors
seqinfo(mi)
sequences(mi)
ranges(mi)
bamfiles(mi)
```

---

ModifierSet-class

*The ModifierSet class*

---

### Description

The ModifierSet class allows multiple [Modifier](#) objects to be created from the same annotation and sequence data varying only the bam input files.

In addition the comparison of samples is also done via calling functions on the ModifierSet objects.

The ModifierSet is a virtual class, which derives from the SimpleList class with the slot elementType = "Modifier". The ModifierSet class has to be implemented for each specific analysis.#'

**Usage**

```

ModifierSet(className, x, annotation, sequences, seqinfo, ...)

## S4 method for signature 'list'
ModifierSet(
  className,
  x,
  annotation = NULL,
  sequences = NULL,
  seqinfo = NULL,
  ...
)

## S4 method for signature 'character'
ModifierSet(
  className,
  x,
  annotation = NULL,
  sequences = NULL,
  seqinfo = NULL,
  ...
)

## S4 method for signature 'BamFileList'
ModifierSet(
  className,
  x,
  annotation = NULL,
  sequences = NULL,
  seqinfo = NULL,
  ...
)

## S4 method for signature 'Modifier'
ModifierSet(className, x, annotation, sequences, seqinfo, ...)

```

**Arguments**

className	The name of the class which should be constructed.
x	the input which can be of the following types <ul style="list-style-type: none"> <li>• Modifier: a single Modifier or a list containing only Modifier objects. The input will just be used as elements of the ModifierSet</li> <li>• BamFileList: a named BamFileList or a list of named BamFileList</li> <li>• list: a list of one or more types of elements: BamFileList, a named list or named character vector. All elements must be or be coercible to a named BamFileList referencing existing bam files. Valid names are control and treated</li> </ul>
annotation	annotation data, which must match the information contained in the BAM files. This parameter is only required, if x is not a Modifier object.
sequences	sequences matching the target sequences the reads were mapped onto. This must match the information contained in the BAM files. This parameter is

only required, if `x` is not a `Modifier` object.

`seqinfo` An optional `Seqinfo` argument or character vector, which can be coerced to one, to subset the sequences to be analyzed on a per chromosome basis.

... Additional optional parameters:

- `internalBP` TRUE or FALSE: should parallelization used internally during creation of each `Modifier` or should the creation of the `Modifier` objects be parallelized? (default: `internalBP = FALSE`). Setting `internalBP` only makes sense, if the `getData` function for `SequenceData` class, the `aggregateData` or the `findMod` function contains parallelized code.

All other arguments will be passed onto the `Modifier` objects.

### Value

a `ModifierSet` object of type `className`

### Creation

The input files have to be provided as a list of elements. Each element in itself must be valid for the creation of `Modifier` object (Have a look at the man page for more details) and must be named.

---

modify

*Searching for modifications in SequenceData*

---

### Description

The `modify` function executes the search for modifications for a `Modifier` class. Usually this is done automatically during construction of a `Modifier` object.

When the `modify` functions is called, the aggregated data is checked for validity for the current settings and the search for modifications is performed using the `findMod`. The results are stored in the modification slot of the `Modifier` object, which is returned by `modify`. The results can be accessed via the `modifications()` function.

`findMod` returns the found modifications as a `GRanges` object and has to be implemented for each individual `Modifier` class.

### Usage

```
modifications(x, ...)
```

```
modify(x, ...)
```

```
findMod(x)
```

```
## S4 method for signature 'Modifier'
modifications(x, perTranscript = FALSE)
```

```
## S4 method for signature 'Modifier'
modify(x, force = FALSE)
```

```
## S4 method for signature 'Modifier'
```

```
findMod(x)

## S4 method for signature 'ModifierSet'
modify(x, force = FALSE)
```

### Arguments

x	a Modifier object.
...	additional arguments
perTranscript	For modifications> TRUE or FALSE: Should the coordinates be returned as local per transcript coordinates?
force	force to run aggregate again, if data is already stored in x.

### Value

- modify: the updated Modifier object.
- modifications: the modifications found as a GRanges object.

### Examples

```
data(msi, package="RNAmoDR")
# modify() triggers the search for modifications in the data contained in
# the Modifier or ModifierSet object
mi <- modify(msi[[1]])
```

---

ModInosine

*ModInosine*


---

### Description

Inosine can be detected in RNA-Seq data by the conversion of A positions to G. This conversion is detected by ModInosine and used to search for Inosine positions. dataType is "PileupSequenceData".

Only samples labeled with the condition treated are used for this analysis, since the A to G conversion is common feature among the reverse transcriptases usually employed. Let us know, if that is not the case, and the class needs to be modified.

Further information on [Functions](#) of ModInosine.

### Usage

```
ModInosine(x, annotation, sequences, seqinfo, ...)
```

```
ModSetInosine(x, annotation = NA, sequences = NA, seqinfo = NA, ...)
```

### Arguments

x	the input which can be of the different types depending on whether a ModRiboMethSeq or a ModSetRiboMethSeq object is to be constructed. For more information have a look at the documentation of the <a href="#">Modifier</a> and <a href="#">ModifierSet</a> classes.
annotation	annotation data, which must match the information contained in the BAM files. This is parameter is only required, if x is not a Modifier object.

sequences	sequences matching the target sequences the reads were mapped onto. This must match the information contained in the BAM files. This parameter is only required, if <code>x</code> is not a <code>Modifier</code> object.
seqinfo	An optional <code>Seqinfo</code> argument or character vector, which can be coerced to one, to subset the sequences to be analyzed on a per chromosome basis.
...	Optional arguments overwriting default values, which are <ul style="list-style-type: none"> <li>• <code>minCoverage</code>: The minimal coverage at the position as integer value (default: <code>minCoverage = 10L</code>).</li> <li>• <code>minReplicate</code>: minimum number of replicates needed for the analysis (default: <code>minReplicate = 1L</code>).</li> <li>• <code>minScore</code>: minimum score to identify Inosine positions de novo (default: <code>minScore = 0.4</code>).</li> </ul>

### Details

ModInosine score: the scores for reported Inosine positions are between 0 and 1. They are calculated as the relative amount of called G bases ( $(G / N)$ ) and only saved for genomic A positions.

### Value

a `ModInosine` or `ModSetInosine` object

### Author(s)

Felix G.M. Ernst [aut]

### Examples

```
# construction of ModInosine object
library(RNAmoR.Data)
library(rtracklayer)
annotation <- GFF3File(RNAmoR.Data.example.man.gff3())
sequences <- RNAmoR.Data.example.man.fasta()
files <- c(treated = RNAmoR.Data.example.wt.1())
mi <- ModInosine(files,annotation = annotation ,sequences = sequences)
# construction of ModSetInosine object
## Not run:
files <- list("SampleSet1" = c(treated = RNAmoR.Data.example.wt.1(),
                             treated = RNAmoR.Data.example.wt.2(),
                             treated = RNAmoR.Data.example.wt.3()),
             "SampleSet2" = c(treated = RNAmoR.Data.example.bud23.1(),
                             treated = RNAmoR.Data.example.bud23.2()),
             "SampleSet3" = c(treated = RNAmoR.Data.example.trm8.1(),
                             treated = RNAmoR.Data.example.trm8.2()))
msi <- ModSetInosine(files, annotation = annotation, sequences = sequences)

## End(Not run)
```

## Description

All of the functions of [Modifier](#) and the [ModifierSet](#) classes are inherited by the `ModInosine` and `ModSetInosine` classes.

Check below for the specifically implemented functions.

## Usage

```
## S4 replacement method for signature 'ModInosine'  
settings(x) <- value  
  
## S4 method for signature 'ModInosine'  
aggregateData(x)  
  
## S4 method for signature 'ModInosine'  
findMod(x)  
  
## S4 method for signature 'ModInosine'  
getDataTrack(x, name, type, ...)  
  
## S4 method for signature 'ModInosine,GRanges'  
plotDataByCoord(x, coord, type = "score", window.size = 15L, ...)  
  
## S4 method for signature 'ModInosine'  
plotData(x, name, from = 1L, to = 30L, type = "score", ...)  
  
## S4 method for signature 'ModSetInosine,GRanges'  
plotDataByCoord(x, coord, type = "score", window.size = 15L, ...)  
  
## S4 method for signature 'ModSetInosine'  
plotData(x, name, from = 1L, to = 30L, type = "score", ...)
```

## Arguments

`x`                    a [Modifier](#) or a [ModifierSet](#) object. For more details see also the man pages for the functions mentioned below.

`value`                See [settings](#)

`coord, name, from, to, type, window.size, ...`  
                      See [plotData](#)

## Details

`ModInosine` specific arguments for [plotData](#):

- `colour.bases` - a named character vector of length = 4 for the colours of the individual bases. The names are expected to be `c("G", "A", "U", "C")`

**Value**

- settings See [settings](#).
- aggregate See [aggregate](#).
- modify See [modify](#).
- getDataTrack a list of [DataTrack](#) objects. See [plotDataByCoord](#).
- plotData See [plotDataByCoord](#).
- plotDataByCoord See [plotDataByCoord](#).

**Examples**

```
data(msi,package="RNAmodR")
mi <- msi[[1]]
settings(mi)
## Not run:
aggregate(mi)
modify(mi)

## End(Not run)
getDataTrack(mi, "1", mainScore(mi))
```

---

ModInosine-internals    *ModInosine internal functions*

---

**Description**

These functions are not intended for general use, but are used for additional package development.

**Arguments**

x, data, seqdata, sequence, args  
internally used arguments

---

NormEndSequenceData-class

*NormEnd5SequenceData/NormEnd3SequenceData*

---

**Description**

The NormEnd5SequenceData/NormEnd3SequenceData aggregate the counts of read ends (Either 5' or 3') at each position along a transcript. In addition, the number of counts are then normalized to the length of the transcript and to the overlapping reads.

Both classes contain three columns per data file named using the following naming convention (normend5/normend3).condition.replicate. The three columns are distinguished by additional identifiers ends, norm.tx and norm.ol.

aggregate calculates the mean and sd for samples in the control and treated condition separately. Similar to the stored results for each of the two conditions six columns are returned (three for mean and sd each) ending in ends, tx and ol.

**Usage**

```

NormEnd5SequenceDataFrame(
  df,
  ranges,
  sequence,
  replicate,
  condition,
  bamfiles,
  seqinfo
)

NormEnd3SequenceDataFrame(
  df,
  ranges,
  sequence,
  replicate,
  condition,
  bamfiles,
  seqinfo
)

NormEnd5SequenceData(bamfiles, annotation, sequences, seqinfo, ...)

NormEnd3SequenceData(bamfiles, annotation, sequences, seqinfo, ...)

## S4 method for signature
## 'NormEnd5SequenceData,BamFileList,GRangesList,XStringSet,ScanBamParam'
getData(x, bamfiles, grl, sequences, param, args)

## S4 method for signature
## 'NormEnd3SequenceData,BamFileList,GRangesList,XStringSet,ScanBamParam'
getData(x, bamfiles, grl, sequences, param, args)

## S4 method for signature 'NormEnd5SequenceData'
aggregateData(x, condition = c("Both", "Treated", "Control"))

## S4 method for signature 'NormEnd3SequenceData'
aggregateData(x, condition = c("Both", "Treated", "Control"))

## S4 method for signature 'NormEnd5SequenceData'
getDataTrack(x, name, ...)

## S4 method for signature 'NormEnd3SequenceData'
getDataTrack(x, name, ...)

```

**Arguments**

df, ranges, sequence, replicate  
 inputs for creating a SequenceDataFrame. See [SequenceDataFrame](#).

condition  
 For [aggregate](#): condition for which the data should be aggregated.

bamfiles, annotation, seqinfo, grl, sequences, param, args, ...  
 See [SequenceData](#) and [SequenceData-functions](#)

x a CoverageSequenceData  
 name For `getDataTrack`: a valid transcript name. Must be a name of ranges(x)

### Value

a NormEnd5SequenceData or NormEnd3SequenceData object

### Examples

```
# Construction of a NormEnd5SequenceData object
## Not run:
library(RNAmoR.Data)
library(rtracklayer)
annotation <- GFF3File(RNAmoR.Data.example.man.gff3())
sequences <- RNAmoR.Data.example.man.fasta()
files <- c(treated = RNAmoR.Data.example.wt.1())
ne5sd <- NormEnd5SequenceData(files, annotation = annotation,
                             sequences = sequences)

## End(Not run)
```

---

PileupSequenceData-class

*PileupSequenceData*

---

### Description

The PileupSequenceData aggregates the pileup of called bases per position.

PileupSequenceData contains five columns per data file named using the following naming convention `pileup.condition.replicate`. The five columns are distinguished by additional identifiers -, G, A, T and C.

`aggregate` calculates the mean and sd for each nucleotide in the control and treated condition separately. The results are then normalized to a row sum of 1.

### Usage

```
PileupSequenceDataFrame(
  df,
  ranges,
  sequence,
  replicate,
  condition,
  bamfiles,
  seqinfo
)

PileupSequenceData(bamfiles, annotation, sequences, seqinfo, ...)

## S4 method for signature
## 'PileupSequenceData,BamFileList,GRangesList,XStringSet,ScanBamParam'
getData(x, bamfiles, grl, sequences, param, args)
```

```
## S4 method for signature 'PileupSequenceData'
aggregateData(x, condition = c("Both", "Treated", "Control"))

## S4 method for signature 'PileupSequenceData'
getDataTrack(x, name, ...)

pileupToCoverage(x)

## S4 method for signature 'PileupSequenceData'
pileupToCoverage(x)
```

### Arguments

df, ranges, sequence, replicate  
                                   inputs for creating a SequenceDataFrame. See [SequenceDataFrame](#).

condition          For [aggregate](#): condition for which the data should be aggregated.

bamfiles, annotation, seqinfo, grl, sequences, param, args, ...  
                                   See [SequenceData](#) and [SequenceData-functions](#)

x                   a PileupSequenceData

name               For [getDataTrack](#): a valid transcript name. Must be a name of ranges(x)

### Value

a PileupSequenceData object

### Examples

```
# Construction of a PileupSequenceData object
library(RNAmoR.Data)
library(rtracklayer)
annotation <- GFF3File(RNAmoR.Data.example.man.gff3())
sequences <- RNAmoR.Data.example.man.fasta()
files <- c(treated = RNAmoR.Data.example.wt.1())
psd <- PileupSequenceData(files, annotation = annotation,
                           sequences = sequences)
```

---

plotData	<i>Visualizing data data from a SequenceData, SequenceDataSet, SequenceDataList, Modifier or ModifierSet object.</i>
----------	--

---

### Description

With the plotData and plotDataByCoord functions data from a SequenceData, SequenceDataSet, SequenceDataList, Modifier or ModifierSet object can be visualized.

Internally the functionality of the Gviz package is used. For each SequenceData and Modifier class the getDataTrack is implemented returning a [DataTrack](#) object from the Gviz package.

Positions to be visualized are selected by defining a genomic coordinate, for which x has to contain data.

**Usage**

```

plotData(x, name, from = 1L, to = 30L, type, ...)

plotDataByCoord(x, coord, type, window.size = 15L, ...)

getDataTrack(x, name, ...)

## S4 method for signature 'Modifier,GRanges'
plotDataByCoord(x, coord, type = NA, window.size = 15L, ...)

## S4 method for signature 'Modifier'
plotData(
  x,
  name,
  from,
  to,
  type = NA,
  showSequenceData = FALSE,
  showSequence = TRUE,
  showAnnotation = FALSE,
  ...
)

## S4 method for signature 'Modifier'
getDataTrack(x, name = name, ...)

## S4 method for signature 'ModifierSet,GRanges'
plotDataByCoord(x, coord, type = NA, window.size = 15L, ...)

## S4 method for signature 'ModifierSet'
plotData(
  x,
  name,
  from,
  to,
  type = NA,
  showSequenceData = FALSE,
  showSequence = TRUE,
  showAnnotation = FALSE,
  ...
)

## S4 method for signature 'SequenceData,GRanges'
plotDataByCoord(x, coord, type = NA, window.size = 15L, ...)

## S4 method for signature 'SequenceData'
plotData(
  x,
  name,
  from,
  to,
  perTranscript = FALSE,

```

```

    showSequence = TRUE,
    showAnnotation = FALSE,
    ...
)

## S4 method for signature 'SequenceData'
getDataTrack(x, name = name, ...)

## S4 method for signature 'SequenceDataList'
getDataTrack(x, name = name, ...)

## S4 method for signature 'SequenceDataList,GRanges'
plotDataByCoord(x, coord, type = NA, window.size = 15L, ...)

## S4 method for signature 'SequenceDataList'
plotData(
  x,
  name,
  from,
  to,
  perTranscript = FALSE,
  showSequence = TRUE,
  showAnnotation = FALSE,
  ...
)

## S4 method for signature 'SequenceDataSet'
getDataTrack(x, name = name, ...)

## S4 method for signature 'SequenceDataSet,GRanges'
plotDataByCoord(x, coord, type = NA, window.size = 15L, ...)

## S4 method for signature 'SequenceDataSet'
plotData(
  x,
  name,
  from,
  to,
  perTranscript = FALSE,
  showSequence = TRUE,
  showAnnotation = FALSE,
  ...
)

```

### Arguments

x	a SequenceData, SequenceDataSet, SequenceDataList, Modifier or ModifierSet object.
name	Only for plotData: the transcript name
from	Only for plotData: start position
to	Only for plotData: end position
type	the data type of data show as data tracks.

...	optional parameters:
	<ul style="list-style-type: none"> <li>• <code>modified.seq</code> TRUE or FALSE. Should the sequence shown with modified nucleotide positions? (default: <code>modified.seq = FALSE</code>)</li> <li>• <code>additional.mod</code> other modifications, which should be shown in the annotation and sequence track. The must be a GRanges compatible with <a href="#">combineIntoModstrings</a>.</li> <li>• <code>annotation.track.pars</code> Parameters passed onto the <a href="#">AnnotationTrack</a>.</li> <li>• <code>sequence.track.pars</code> Parameters passed onto the <a href="#">SequenceTrack</a>.</li> </ul>
<code>coord</code>	coordinates of a positions to subset to as a GRanges object. The 'Parent' column is expected to match the transcript name.
<code>window.size</code>	integer value for the number of positions on the left and right site of the selected positions included in the plotting (default: <code>window.size = 15L</code> )
<code>showSequenceData</code>	TRUE or FALSE: should the sequence data be shown? (default: <code>seqdata = FALSE</code> )
<code>showSequence</code>	TRUE or FALSE: should a sequence track be shown? (default: <code>seqdata = TRUE</code> )
<code>showAnnotation</code>	TRUE or FALSE: should a annotation track be shown? (default: <code>seqdata = FALSE</code> )
<code>perTranscript</code>	TRUE or FALSE: Should the positions shown per transcript? (default: <code>perTranscript = FALSE</code> )

**Value**

a plot send to the active graphic device

**Examples**

```
data(msi,package="RNAmodR")
plotData(msi[[1]], "2", from = 10L, to = 45L)
## Not run:
plotData(msi, "2", from = 10L, to = 45L)

## End(Not run)
```

---

plotROC

*ROCR functions for Modifier and ModifierSet objects*

---

**Description**

plotROC streamlines labeling, prediction, performance and plotting functions to test the performance of a Modifier object and the data analyzed via the functionality from the ROCR package.

The data from `x` will be labeled as positive using the `coord` arguments. The other arguments will be passed on to the specific ROCR functions.

By default the `prediction.args` include three values:

- `measure = "tpr"`
- `x.measure = "fpr"`
- `score = mainScore(x)`

The remaining arguments are not predefined.

**Usage**

```
plotROC(x, coord, ...)  
  
## S4 method for signature 'Modifier'  
plotROC(  
  x,  
  coord,  
  score = NULL,  
  prediction.args = list(),  
  performance.args = list(),  
  plot.args = list()  
)  
  
## S4 method for signature 'ModifierSet'  
plotROC(  
  x,  
  coord,  
  score = NULL,  
  prediction.args = list(),  
  performance.args = list(),  
  plot.args = list()  
)
```

**Arguments**

x	a Modifier or a ModifierSet object
coord	coordinates of position to label as positive. Either a GRanges or a GRangesList object. For both types the Parent column is expected to match the gene or transcript name.
...	additional arguments
score	the score identifier to subset to, if multiple scores are available.
prediction.args	arguments which will be used for calling <code>prediction</code> form the ROCR package
performance.args	arguments which will be used for calling <code>performance</code> form the ROCR package
plot.args	arguments which will be used for calling <code>plot</code> on the performance object of the ROCR package. If multiple scores are plotted (for example if the score argument is not explicitly set) <code>add = FALSE</code> will be set.

**Value**

a plot send to the active graphic device

**References**

Tobias Sing, Oliver Sander, Niko Beerenwinkel, Thomas Lengauer (2005): "ROCR: visualizing classifier performance in R." *Bioinformatics* 21(20):3940-3941 DOI: [10.1093/bioinformatics/bti623](https://doi.org/10.1093/bioinformatics/bti623)

**Examples**

```

data(msi,package="RNAmodR")
# constructing a GRanges object to mark positive positions
mod <- modifications(msi)
coord <- unique(unlist(mod))
coord$score <- NULL
coord$sd <- NULL
# plotting a TPR vs. FPR plot per ModInosine object
plotROC(msi[[1]],coord)
# plotting a TPR vs. FPR plot per ModSetInosine object
plotROC(msi,coord)

```

---

ProtectedEndSequenceData-class

*ProtectedEndSequenceData*


---

**Description**

ProtectedEndSequenceData implements [SequenceData](#) to contain and aggregate the start and ends of reads per position along a transcript. ProtectedEndSequenceData offsets the start position by -1 to align the information on the 5'-3'-phosphate bonds to one position. The ProtectedEndSequenceData class is implemented specifically as required for the RiboMethSeq method.

The objects of type ProtectedEndSequenceData contain three columns per data file named using the following naming convention protectedend.condition.replicate.

aggregate calculates the mean and sd for samples in the control and treated condition separately.

**Usage**

```

ProtectedEndSequenceDataFrame(
  df,
  ranges,
  sequence,
  replicate,
  condition,
  bamfiles,
  seqinfo
)

```

```

ProtectedEndSequenceData(bamfiles, annotation, sequences, seqinfo, ...)

```

```

## S4 method for signature
## 'ProtectedEndSequenceData,
## BamFileList,
## GRangesList,
## XStringSet,
## ScanBamParam'
getData(x, bamfiles, grl, sequences, param, args)

```

```

## S4 method for signature 'ProtectedEndSequenceData'
aggregateData(x, condition = c("Both", "Treated", "Control"))

```

```
## S4 method for signature 'ProtectedEndSequenceData'
getDataTrack(x, name, ...)
```

### Arguments

df, ranges, sequence, replicate  
                                   inputs for creating a SequenceDataFrame. See [SequenceDataFrame](#).

condition          For [aggregate](#): condition for which the data should be aggregated.

bamfiles, annotation, seqinfo, grl, sequences, param, args, ...  
                                   See [SequenceData](#) and [SequenceData-functions](#)

x                   a ProtectedEndSequenceData

name               For [getDataTrack](#): a valid transcript name. Must be a name of ranges(x)

### Value

a ProtectedEndSequenceData object

### Examples

```
# Construction of a ProtectedEndSequenceData object
library(RNAmodR.Data)
library(rtracklayer)
annotation <- GFF3File(RNAmodR.Data.example.man.gff3())
sequences <- RNAmodR.Data.example.man.fasta()
files <- c(treated = RNAmodR.Data.example.wt.1())
pesd <- ProtectedEndSequenceData(files, annotation = annotation,
                                sequences = sequences)
```

---

RNAmodR

*RNAmodR*

---

### Description

Post-transcriptional modifications can be found abundantly in rRNA and tRNA and can be detected classically via several strategies. However, difficulties arise if the identity and the position of the modified nucleotides is to be determined at the same time. Classically, a primer extension, a form of reverse transcription (RT), would allow certain modifications to be accessed by blocks during the RT changes or changes in the cDNA sequences. Other modification would need to be selectively treated by chemical reactions to influence the outcome of the reverse transcription.

With the increased availability of high throughput sequencing, these classical methods were adapted to high throughput methods allowing more RNA molecules to be accessed at the same time. With these advances post-transcriptional modifications were also detected on mRNA. Among these high throughput techniques are for example Pseudo-Seq (Carlile et al. 2014), RiboMethSeq (Birkedal et al. 2015) and AlkAnilineSeq (Marchand et al. 2018) each able to detect a specific type of modification from footprints in RNA-Seq data prepared with the selected methods.

Since similar pattern can be observed from some of these techniques, overlaps of the bioinformatical pipeline already are and will become more frequent with new emerging sequencing techniques.

RNAmodR implements classes and a workflow to detect post-transcriptional RNA modifications in high throughput sequencing data. It is easily adaptable to new methods and can help during the phase of initial method development as well as more complex screenings.

Briefly, from the `SequenceData`, specific subclasses are derived for accessing specific aspects of aligned reads, e.g. 5'-end positions or pileup data. With this a `Modifier` class can be used to detect specific patterns for individual types of modifications. The `SequenceData` classes can be shared by different `Modifier` classes allowing easy adaptation to new methods.

### Author(s)

Felix G M Ernst [aut], Denis L.J. Lafontaine [ctb]

### References

- Carlile TM, Rojas-Duran MF, Zinshteyn B, Shin H, Bartoli KM, Gilbert WV (2014): "Pseudouridine profiling reveals regulated mRNA pseudouridylation in yeast and human cells." *Nature* 515 (7525), P. 143–146. DOI: [10.1038/nature13802](https://doi.org/10.1038/nature13802).
- Birkedal U, Christensen-Dalsgaard M, Krogh N, Sabarinathan R, Gorodkin J, Nielsen H (2015): "Profiling of ribose methylations in RNA by high-throughput sequencing." *Angewandte Chemie (International ed. in English)* 54 (2), P. 451–455. DOI: [10.1002/anie.201408362](https://doi.org/10.1002/anie.201408362).
- Marchand V, Ayadi L, \_\_Ernst FGM\_\_, Hertler J, Bourguignon-Igel V, Galvanin A, Kotter A, Helm M, \_\_Lafontaine DLJ\_\_, Motorin Y (2018): "AlkAniline-Seq: Profiling of m7 G and m3 C RNA Modifications at Single Nucleotide Resolution." *Angewandte Chemie (International ed. in English)* 57 (51), P. 16785–16790. DOI: [10.1002/anie.201810946](https://doi.org/10.1002/anie.201810946).

### See Also

The `RNAmoDR.RiboMethSeq` and `RNAmoDR.AlkAnilineSeq` package.

---

RNAmoDR-datasets

*Example data in the RNAmoDR package*

---

### Description

The following datasets are contained in the `RNAmoDR` package. They are used in the man page examples.

### Usage

`data(msi)`

`data(sds)`

`data(sd1)`

`data(psd)`

`data(e5sd)`

`data(e3sd)`

`data(esd)`

`data(csd)`

```
data(ne3sd)
```

```
data(ne5sd)
```

```
data(pesd)
```

**Format**

- msi a ModSetInosine instance
- sds a SequenceDataSet instance
- sdl a SequenceDataList instance
- psd a PileupSequenceData instance
- e5sd a End5SequenceData instance
- e3sd a End3SequenceData instance
- esd a EndSequenceData instance
- csd a CoverageSequenceData instance
- ne3sd a NormEnd3SequenceData instance
- ne5sd a NormEnd5SequenceData instance
- pesd a ProtectedEndSequenceData instance

An object of class SequenceDataSet of length 2.

An object of class SequenceDataList of length 3.

An object of class PileupSequenceData of dimension 100 x 101 x 15 x 15.

An object of class End5SequenceData of dimension 100 x 101 x 3 x 3.

An object of class End3SequenceData of dimension 100 x 101 x 3 x 3.

An object of class EndSequenceData of dimension 100 x 101 x 3 x 3.

An object of class CoverageSequenceData of dimension 100 x 101 x 3 x 3.

An object of class NormEnd3SequenceData of dimension 100 x 101 x 9 x 9.

An object of class NormEnd5SequenceData of dimension 100 x 101 x 9 x 9.

An object of class ProtectedEndSequenceData of dimension 100 x 101 x 3 x 3.

---

RNAmoDR-development    *RNAmoDR developments functions*

---

**Description**

These functions are not intended for general use, but are used for additional package development.

getData is used to load data into a [SequenceData](#) object and must be implemented for all SequenceData classes. The results must match the requirements outlined in the value section.

In addition the following functions should be implemented for complete functionality:

aggregateData for each SequenceData and Modifier class. See also [aggregateData](#)

findMod for each Modifier class. See also [findMod](#).

plotData/plotDataByCoord for each Modifier and ModifierSet class. See also [plotData](#).

The following helper function can be called from within `findMod` to construct a coordinate for each modification found:

`constructModRanges` constructs a `GRanges` object describing the location, type and associated scores of a modification. `constructModRanges` is typically called from the `modify` function, which must be implemented for all `Modifier` classes.

### Usage

```
constructModRanges(range, data, modType, scoreFun, source, type)
```

```
getData(x, bamfiles, grl, sequences, param, args)
```

```
## S4 method for signature 'GRanges,DataFrame'
```

```
constructModRanges(range, data, modType, scoreFun, source, type)
```

### Arguments

<code>range</code>	for <code>constructModRanges</code> : a <code>GRanges</code> object
<code>data</code>	for <code>constructModRanges</code> : a <code>DataFrame</code> object
<code>modType</code>	for <code>constructModRanges</code> : a valid <code>shortName</code> for the modification found. Must be present in <code>shortName(ModRNAString())</code> .
<code>scoreFun</code>	for <code>constructModRanges</code> : a custom function for extracting scores from data. The result must be a list.
<code>source</code>	for <code>constructModRanges</code> : a single character vector for populating the source column of the result.
<code>type</code>	for <code>constructModRanges</code> : a single character vector for populating the source column of the result.
<code>x</code>	for <code>getData</code> : a <code>SequenceData</code> object.
<code>bamfiles</code>	for <code>getData</code> : a <code>BamFileList</code> object.
<code>grl</code>	for <code>getData</code> : a <code>GRangesList</code> object.
<code>sequences</code>	for <code>getData</code> : a <code>XStringSet</code> object.
<code>param</code>	for <code>getData</code> : a <code>ScanBamParam</code> object.
<code>args</code>	for <code>getData</code> : a list with optional arguments.

### Value

- `getData`: returns a list with elements per `BamFile` in `bamfiles`. Elements can be `IntegerList`, `NumericList` or a `CompressedSplitDataFrameList`. The data in the elements must be order by increasing positions numbers. However, names and rownames will be discarded.
- `constructModRanges`: returns a `GRanges` object with genomic coordinates of modified nucleotides in the associated transcripts.

### Examples

```
# new SequenceData class
setClass(Class = "ExampleSequenceData",
         contains = "SequenceData",
         prototype = list(minQuality = 5L))
ExampleSequenceData <- function(bamfiles, annotation, sequences, seqinfo, ...){
  RNAmodR:::SequenceData("Example", bamfiles = bamfiles,
```

```

        annotation = annotation, sequences = sequences,
        seqinfo = seqinfo, ...)
    }
    setMethod("getData",
              signature = c(x = "ExampleSequenceData",
                           bamfiles = "BamFileList",
                           grl = "GRangesList",
                           sequences = "XStringSet",
                           param = "ScanBamParam"),
              definition = function(x, bamfiles, grl, sequences, param, args){
                ###
              }
    )
    setMethod("aggregateData",
              signature = c(x = "ExampleSequenceData"),
              function(x, condition = c("Both", "Treated", "Control")){
                ###
              }
    )
    setMethod(
      f = "getDataTrack",
      signature = c(x = "ExampleSequenceData"),
      definition = function(x, name, ...) {
        ###
      }
    )
    )

# new Modifier class
setClass("ModExample",
         contains = "Modifier",
         prototype = list(mod = "X",
                          score = "score",
                          dataType = "ExampleSequenceData"))
ModExample <- function(x, annotation, sequences, seqinfo, ...){
  RNAmodR::Modifier("ModExample", x = x, annotation = annotation,
                    sequences = sequences, seqinfo = seqinfo, ...)
}

setMethod(f = "aggregateData",
          signature = c(x = "ModExample"),
          definition =
            function(x, force = FALSE){
              # Some data with element per transcript
            }
          )

setMethod("findMod",
          signature = c(x = "ModExample"),
          function(x){
            # an element per modification found.
          }
          )

setMethod(
  f = "getDataTrack",
  signature = signature(x = "ModExample"),
  definition = function(x, name, type, ...) {
  }
)

```

```

)
setMethod(
  f = "plotDataByCoord",
  signature = signature(x = "ModExample", coord = "GRanges"),
  definition = function(x, coord, type = "score", window.size = 15L, ...) {
  }
)
setMethod(
  f = "plotData",
  signature = signature(x = "ModExample"),
  definition = function(x, name, from, to, type = "score", ...) {
  }
)

# new ModifierSet class
setClass("ModSetExample",
  contains = "ModifierSet",
  prototype = list(elementType = "ModExample"))
ModSetExample <- function(x, annotation, sequences, seqinfo, ...){
  RNAmodR:::ModifierSet("ModExample", x = x, annotation = annotation,
    sequences = sequences, seqinfo = seqinfo, ...)
}

setMethod(
  f = "plotDataByCoord",
  signature = signature(x = "ModSetExample", coord = "GRanges"),
  definition = function(x, coord, type = "score", window.size = 15L, ...) {
  }
)
setMethod(
  f = "plotData",
  signature = signature(x = "ModSetExample"),
  definition = function(x, name, from, to, type = "score", ...) {
  }
)

```

---

 RNAmodR-internals

*RNAmodR internal functions*


---

## Description

These functions are used internally.

## Usage

```
## S4 method for signature 'SequenceDataSet'
parallel_slot_names(x)
```

```
## S4 method for signature 'SequenceDataSet'
getListElement(x, i, exact = TRUE)
```

```
## S4 method for signature 'SequenceDataList'
parallel_slot_names(x)
```

```
## S4 method for signature 'SequenceDataList'
getListElement(x, i, exact = TRUE)

## S4 method for signature 'Modifier'
subset(x, name, pos = 1L, ...)

## S4 method for signature 'SplitDataFrameList,GRanges'
labelByCoord(x, coord, ...)
```

### Arguments

object, range, data, modType, scoreFun, source, type, x, i, j, ..., exact,  
value

Internal arguments

### Value

internally used values

---

SequenceData-class      *The SequenceData class*

---

### Description

The SequenceData class is implemented to contain data on each position along transcripts and holds the corresponding annotation data and nucleotide sequence of these transcripts. To access this data several [functions](#) are available. The SequenceData class is a virtual class, from which specific classes can be extended. Currently the following classes are implemented:

- [CoverageSequenceData](#)
- [End5SequenceData](#), [End3SequenceData](#), [EndSequenceData](#)
- [NormEnd5SequenceData](#), [NormEnd5SequenceData](#)
- [PileupSequenceData](#)
- [ProtectedEndSequenceData](#)

The annotation and sequence data can be accessed through the functions `ranges` and `sequences`, respectively. Be aware, that the data is always provided according to genomic positions with increasing rownames, but the sequence is given as the actual sequence of the transcript. Therefore, it is necessary to treat the minus strand accordingly.

The SequenceData class is derived from the [CompressedSplitDataFrameList](#) class with additional slots for annotation and sequence data. Some functionality is not inherited and might not be available to full extend, e.g. `relist`.

#### SequenceDataFrame

The SequenceDataFrame class is a virtual class and contains data for positions along a single transcript. In addition to being used for returning elements from a SequenceData object, the SequenceDataFrame class is used to store the unlisted data within a [SequenceData](#) object. Therefore, a matching SequenceData and SequenceDataFrame class must be implemented.

The SequenceDataFrame class is derived from the [DataFrame](#) class.

Subsetting of a SequenceDataFrame returns a SequenceDataFrame or DataFrame, if it is subset by a column or row, respectively. The `drop` argument is ignored for column subsetting.

**Usage**

```

## S4 method for signature 'SequenceData'
cbind(..., deparse.level = 1)

## S4 method for signature 'SequenceData'
rbind(..., deparse.level = 1)

SequenceData(dataType, bamfiles, annotation, sequences, seqinfo, ...)

## S4 method for signature 'character,character'
SequenceData(dataType, bamfiles, annotation, sequences, seqinfo, ...)

## S4 method for signature 'character,BSgenome'
SequenceData(dataType, bamfiles, annotation, sequences, seqinfo, ...)

## S4 method for signature 'TxDb,character'
SequenceData(dataType, bamfiles, annotation, sequences, seqinfo, ...)

## S4 method for signature 'TxDb,BSgenome'
SequenceData(dataType, bamfiles, annotation, sequences, seqinfo, ...)

## S4 method for signature 'GRangesList,character'
SequenceData(dataType, bamfiles, annotation, sequences, seqinfo, ...)

## S4 method for signature 'GRangesList,BSgenome'
SequenceData(dataType, bamfiles, annotation, sequences, seqinfo, ...)

## S4 method for signature 'GFF3File,BSgenome'
SequenceData(dataType, bamfiles, annotation, sequences, seqinfo, ...)

## S4 method for signature 'GFF3File,character'
SequenceData(dataType, bamfiles, annotation, sequences, seqinfo, ...)

## S4 method for signature 'character,FaFile'
SequenceData(dataType, bamfiles, annotation, sequences, seqinfo, ...)

## S4 method for signature 'GFF3File,FaFile'
SequenceData(dataType, bamfiles, annotation, sequences, seqinfo, ...)

## S4 method for signature 'TxDb,FaFile'
SequenceData(dataType, bamfiles, annotation, sequences, seqinfo, ...)

## S4 method for signature 'GRangesList,FaFile'
SequenceData(dataType, bamfiles, annotation, sequences, seqinfo, ...)

```

**Arguments**

- ... Optional arguments overwriting default values. Not all SequenceData classes use all arguments. The arguments are:
- `minLength` single integer value setting a threshold for minimum read length. Shorter reads are discarded (default: `minLength = NA`).

- `maxLength` single integer value setting a threshold for maximum read length. Longer reads are discarded (default: `maxLength = NA`).
- `minQuality` single integer value setting a threshold for maximum read quality. Reads with a lower quality are discarded (default: `minQuality = 5L`, but this is class dependent).
- `max_depth` maximum depth for pileup loading (default: `max_depth = 10000L`).

<code>deparse.level</code>	See <a href="#">base::cbind</a> for a description of this argument.
<code>dataType</code>	The prefix for construction the class name of the <code>SequenceData</code> subclass to be constructed.
<code>bamfiles</code>	the input which can be of the following types <ul style="list-style-type: none"> <li>• <code>BamFileList</code>: a named <code>BamFileList</code></li> <li>• <code>character</code>: a character vector, which must be coercible to a named <code>BamFileList</code> referencing existing bam files. Valid names are <code>control</code> and <code>replicates</code> to define conditions and replicates</li> </ul>
<code>annotation</code>	annotation data, which must match the information contained in the BAM files.
<code>sequences</code>	sequences matching the target sequences the reads were mapped onto. This must match the information contained in the BAM files.
<code>seqinfo</code>	optional <a href="#">Seqinfo</a> to subset the transcripts analyzed on a chromosome basis.

**Value**

A `SequenceData` object

**Slots**

`sequencesType` a character value for the class name of sequences. Either `RNAStringSet`, `ModRNAStringSet`, `DNAStringSet` or `ModDNAStringSet`.

`minQuality` a integer value describing a threshold of the minimum quality of reads to be used.

---

SequenceData-functions

*SequenceData/SequenceDataSet/SequenceDataList/SequenceDataFrame  
functions*

---

**Description**

The `SequenceData`, `SequenceDataSet`, `SequenceDataList` and `SequenceDataFrame` classes share functionality. Have a look at the elements listed directly below.

**Usage**

```
replicates(x)
```

```
## S4 method for signature 'SequenceDataFrame'  
show(object)
```

```
## S4 method for signature 'SequenceDataFrame'  
conditions(object)
```

```
## S4 method for signature 'SequenceDataFrame'  
bamfiles(x)  
  
## S4 method for signature 'SequenceDataFrame'  
dataType(x)  
  
## S4 method for signature 'SequenceDataFrame'  
ranges(x)  
  
## S4 method for signature 'SequenceDataFrame'  
replicates(x)  
  
## S4 method for signature 'SequenceDataFrame'  
seqinfo(x)  
  
## S4 method for signature 'SequenceDataFrame'  
seqinfo(x)  
  
## S4 method for signature 'SequenceDataFrame'  
seqtype(x)  
  
## S4 replacement method for signature 'SequenceDataFrame'  
seqtype(x) <- value  
  
## S4 method for signature 'SequenceDataFrame'  
sequences(x)  
  
## S4 method for signature 'SequenceData'  
show(object)  
  
## S4 method for signature  
## 'SequenceData,BamFileList,GRangesList,XStringSet,ScanBamParam'  
getData(x, bamfiles, grl, sequences, param, args)  
  
## S4 method for signature 'SequenceData'  
bamfiles(x)  
  
## S4 method for signature 'SequenceData'  
conditions(object)  
  
## S4 method for signature 'SequenceData'  
ranges(x)  
  
## S4 method for signature 'SequenceData'  
replicates(x)  
  
## S4 method for signature 'SequenceData'  
seqinfo(x)  
  
## S4 method for signature 'SequenceData'  
sequences(x)
```

```
## S4 method for signature 'SequenceData'
seqtype(x)

## S4 replacement method for signature 'SequenceData'
seqtype(x) <- value

## S4 method for signature 'SequenceData'
dataType(x)

## S4 method for signature 'SequenceDataSet'
show(object)

## S4 method for signature 'SequenceDataSet'
bamfiles(x)

## S4 method for signature 'SequenceDataSet'
conditions(object)

## S4 method for signature 'SequenceDataSet'
names(x)

## S4 method for signature 'SequenceDataSet'
ranges(x)

## S4 method for signature 'SequenceDataSet'
replicates(x)

## S4 method for signature 'SequenceDataSet'
seqinfo(x)

## S4 method for signature 'SequenceDataSet'
seqtype(x)

## S4 replacement method for signature 'SequenceDataSet'
seqtype(x) <- value

## S4 method for signature 'SequenceDataSet'
sequences(x)

## S4 method for signature 'SequenceDataList'
show(object)

## S4 method for signature 'SequenceDataList'
bamfiles(x)

## S4 method for signature 'SequenceDataList'
conditions(object)

## S4 method for signature 'SequenceDataList'
names(x)
```

```

## S4 method for signature 'SequenceDataList'
ranges(x)

## S4 method for signature 'SequenceDataList'
replicates(x)

## S4 method for signature 'SequenceDataList'
seqinfo(x)

## S4 method for signature 'SequenceDataList'
seqtype(x)

## S4 replacement method for signature 'SequenceDataList'
seqtype(x) <- value

## S4 method for signature 'SequenceDataList'
sequences(x)

```

### Arguments

<code>x, object</code>	a <code>SequenceData</code> , <code>SequenceDataSet</code> , <code>SequenceDataList</code> or a <code>SequenceDataFrame</code> object.
<code>value</code>	a new <code>seqtype</code> , either "RNA" or "DNA"
<code>bamfiles</code>	a <code>BamFileList</code> .
<code>grl</code>	a <code>GRangesList</code> from <code>exonsBy(..., by = "tx")</code>
<code>sequences</code>	a <code>XStringSet</code> of type <code>RNAStringSet</code> , <code>ModRNAStringSet</code> , <code>DNAStringSet</code> or <code>ModDNAStringSet</code>
<code>param</code>	a <code>ScanBamParam</code> object
<code>args</code>	a list of addition arguments

### Value

- `seqinfo`: a `Seqinfo` object (`()`).
- `sequences`: a `RNAstingSet` object or a `RNAString` object for a `SequenceDataFrame`.
- `ranges`: a `GRangesList` object with each element per transcript or a `GRanges` object for a `SequenceDataFrame`.
- `bamfiles`: a `BamFileList` object or a `SimpleList` of `BamFileList` objects for a `SequenceDataList`.

### Examples

```

data(e5sd, package="RNAmoR")
# general accessors
seqinfo(e5sd)
sequences(e5sd)
ranges(e5sd)
bamfiles(e5sd)

```

---

SequenceDataFrame-class

*The SequenceDataFrame class*


---

## Description

The SequenceDataFrame class is a virtual class and contains data for positions along a single transcript. In addition to being used for returning elements from a SequenceData object, the SequenceDataFrame class is used to store the unlisted data within a SequenceData object. Therefore, a matching SequenceData and SequenceDataFrame class must be implemented.

The SequenceDataFrame class is derived from the DataFrame class. To follow the functionality in the S4Vectors package, SequenceDataFrame implements the concept, whereas SequenceDataFrame is the implementation for in-memory data representation from which some specific \*SequenceDataFrame class derive from, e.g. CoverageSequenceData.

Subsetting of a SequenceDataFrame returns a SequenceDataFrame or DataFrame, if it is subset by a column or row, respectively. The drop argument is ignored for column subsetting.

## Usage

```
## S4 method for signature 'SequenceDataFrame'
cbind(..., deparse.level = 1)

## S4 method for signature 'SequenceDataFrame,ANY,ANY,ANY'
x[i, j, ..., drop = TRUE]
```

## Arguments

x, i, j, ..., drop, deparse.level  
arguments used for [subsetting](#) or [base::cbind](#).

## Value

A SequenceDataFrame object or if subset to row a DataFrame

## Slots

ranges a GRanges object each element describing a transcript including its element. The GRanges is constructed from the unlisted results of the `exonsBy(x, by="tx")` function. If during construction a GRangesList is provided instead of a character value pointing to a gff3 file or a TxDb object, it must have a comparable structure.

sequence a XString of type sequencesType from the parent SequenceData object.

condition conditions along the BamFileList: Either control or treated

replicate replicate number along the BamFileList for each of the condition types.

bamfiles the input bam files as BamFileList

seqinfo a Seqinfo describing the available/used chromosomes.

## See Also

for an example see [ProtectedEndSequenceData](#) and for more information see [SequenceData](#)

**Examples**

```

data(e5sd,package="RNAmodR")
# A SequenceDataFrame can is usually constructed by subsetting from
# a SequenceData object
sdf <- e5sd[[1]]
# Its also used to store the unlisted data in a SequenceData object
sdf <- unlist(e5sd) # should probably only used internally
e5sd <- relist(sdf,e5sd)

```

---

**SequenceDataList-class***The SequenceDataList class*

---

**Description**

The SequenceDataList class is used to hold SequenceData or SequenceDataSet objects as its elements. It is derived from the [List](#) class.

The SequenceDataList is used to hold data from different sets of aligned reads. This allows multiple methods to be aggregated into one modification detection strategy. Annotation and sequence data must be the same for all elements, however the bam files can be different.

**Usage**

```
SequenceDataList(...)
```

**Arguments**

... The elements to be included in the SequenceDataList.

**Value**

a SequenceDataList

**Examples**

```

data(psd,package="RNAmodR")
data(e5sd,package="RNAmodR")
sdl <- SequenceDataList(SequenceDataSet(psd,e5sd),e5sd)

```

---

**SequenceDataSet-class** *The SequenceDataSet class*

---

**Description**

The SequenceDataSet class is used to hold SequenceData objects as its elements. It is derived from the [List](#) class.

The SequenceDataSet is used to hold different data types from the of same aligned reads. The same dataset can be used to generate multiple sets of data types. Bam files, annotation and sequence data must be the same for all elements.

**Usage**

```
SequenceDataSet(...)
```

**Arguments**

... The elements to be included in the SequenceDataSet.

**Value**

a SequenceDataSet

**Examples**

```
data(psd, package="RNAmodR")
data(e5sd, package="RNAmodR")
sd1 <- SequenceDataSet(psd, e5sd)
```

---

SequenceModDNAStringSetTrack-class

*ModDNASequenceTrack*

---

**Description**

A Gviz compatible [SequenceTrack](#) for showing modified DNA sequences.

**Usage**

```
ModDNASequenceTrack(sequence, chromosome, genome, name = "SequenceTrack", ...)
```

```
## S4 method for signature 'SequenceModDNAStringSetTrack'
seqnames(x)
```

```
## S4 method for signature 'SequenceModDNAStringSetTrack'
seqlevels(x)
```

**Arguments**

sequence A character vector or ModDNAString object of length one. The sequence to display. See [SequenceTrack](#).

chromosome, genome, name, ...  
See [SequenceTrack](#).

x A SequenceModDNAStringSetTrack object.

**Value**

a SequenceModDNAStringSetTrack object

**Slots**

sequence A ModDNAStringSet object

**Examples**

```
seq <- ModDNAStringSet(c(chr1 = paste0(alphabet(ModDNAString()),
                                       collapse = "")))
st <- ModDNASequenceTrack(seq)
Gviz::plotTracks(st, chromosome = "chr1", from = 1L, to = 20L)
```

---

SequenceModRNAStringSetTrack-class  
*ModRNASequenceTrack*

---

**Description**

A Gviz compatible [SequenceTrack](#) for showing modified RNA sequences.

**Usage**

```
ModRNASequenceTrack(sequence, chromosome, genome, name = "SequenceTrack", ...)
```

```
## S4 method for signature 'SequenceModRNAStringSetTrack'
seqnames(x)
```

```
## S4 method for signature 'SequenceModRNAStringSetTrack'
seqlevels(x)
```

**Arguments**

**sequence** A character vector or `ModRNAString` object of length one. The sequence to display. See [SequenceTrack](#).

**chromosome, genome, name, ...** See [SequenceTrack](#).

**x** A `SequenceModRNAStringSetTrack` object.

**Value**

a `SequenceModRNAStringSetTrack` object

**Slots**

**sequence** A `ModRNAStringSet` object

**Examples**

```
seq <- ModRNAStringSet(c(chr1 = paste0(alphabet(ModRNAString()),
                                       collapse = "")))
st <- ModRNASequenceTrack(seq)
Gviz::plotTracks(st, chromosome = "chr1", from = 1L, to = 20L)
```

---

settings	<i>Settings for Modifier objects</i>
----------	--------------------------------------

---

**Description**

Depending on data preparation, quality and desired stringency of a modification strategy, settings for cut off parameters or other variables may need to be adjusted. This should be rarely the case, but a function for changing these settings, is implemented as the... settings function.

For changing values the input can be either a list or something coercible to a list. Upon changing a setting, the validity of the value in terms of type(!) and dimensions will be checked.

If settings have been modified after the data was loaded, the data is potentially invalid. To update the data, run the aggregate or the modify function.

**Usage**

```
settings(x, name = NULL)

settings(x, name) <- value

## S4 method for signature 'Modifier'
settings(x, name = NULL)

## S4 replacement method for signature 'Modifier'
settings(x) <- value

## S4 method for signature 'ModifierSet'
settings(x, name = NULL)

## S4 replacement method for signature 'ModifierSet'
settings(x) <- value
```

**Arguments**

x	a Modifier or ModifierSet class
name	name of the setting to be returned or set
value	value of the setting to be set

**Value**

If name is omitted, settings returns a list of all settings. If name is set, settings returns a single settings or NULL, if a value for name is not available.

**Examples**

```
data(msi, package="RNAmodR")
mi <- msi[[1]]
# returns a list of all settings
settings(mi)
# accesses a specific setting
settings(mi, "minCoverage")
# modification of setting
settings(mi) <- list(minCoverage = 11L)
```

stats

*Retrieving information about used reads in RNAmoDR***Description**

stats returns information about reads used in the RNAmoDR analysis. Three modes are available depending on which type of object is provided. If a [SequenceData](#) object is provided, a [BamFile](#) or [BamFileList](#) must be provided as well. If a [Modifier](#) object is used, the bam files returned from the bamfiles function are used. This is also the case, if a [ModifierSet](#) object is used.

**Usage**

```
stats(x, file, ...)

## S4 method for signature 'SequenceData,BamFile'
stats(x, file, ...)

## S4 method for signature 'SequenceData,BamFileList'
stats(x, file, ...)

## S4 method for signature 'Modifier,missing'
stats(x)

## S4 method for signature 'ModifierSet,missing'
stats(x)
```

**Arguments**

x a [SequenceData](#), [Modifier](#) or [ModifierSet](#) object  
file a [BamFile](#) or [BamFileList](#), if x is a [SequenceData](#) object.  
... optional parameters used as stated [here](#) (except minQuality), if x is a [SequenceData](#) object.

**Value**

a [DataFrame](#), [DataFrameList](#) or [SimpleList](#) with the results in aggregated form

**Examples**

```
library(RNAmoDR.Data)
library(rtracklayer)
sequences <- RNAmoDR.Data.example.AAS.fasta()
annotation <- GFF3File(RNAmoDR.Data.example.AAS.gff3())
files <- list("SampleSet1" = c(treated = RNAmoDR.Data.example.wt.1(),
                             treated = RNAmoDR.Data.example.wt.2(),
                             treated = RNAmoDR.Data.example.wt.3()),
            "SampleSet2" = c(treated = RNAmoDR.Data.example.bud23.1(),
                             treated = RNAmoDR.Data.example.bud23.2()),
            "SampleSet3" = c(treated = RNAmoDR.Data.example.trm8.1(),
                             treated = RNAmoDR.Data.example.trm8.2()))
msi <- ModSetInosine(files, annotation = annotation, sequences = sequences)
# smallest chunk of information
```

```

stats(sequenceData(msi[[1L]]),bamfiles(msi[[1L]])[[1L]])
# partial information
stats(sequenceData(msi[[1L]]),bamfiles(msi[[1L]]))
# the whole stats
stats(msi)

```

---

subsetByCoord	<i>Subsetting data from a SequenceData, SequenceDataSet, SequenceDataList, Modifier or ModifierSet object.</i>
---------------	--

---

## Description

With the subsetByCoord function data from a SequenceData, SequenceDataSet, SequenceDataList, Modifier or ModifierSet object can be subset to positions as defined in coord.

If coord contains a column mod and x is a Modifier object, it will be filtered to identifiers matching the `modType` of x. To disable this behaviour remove the column mod from coord or set type = NA

labelByCoord functions similarly. It will return a SplitDataFrameList, which matches the dimensions of the aggregated data plus the labels column, which contains logical values to indicate selected positions.

## Usage

```

subsetByCoord(x, coord, ...)

labelByCoord(x, coord, ...)

## S4 method for signature 'Modifier,GRanges'
subsetByCoord(x, coord, ...)

## S4 method for signature 'Modifier,GRangesList'
subsetByCoord(x, coord, ...)

## S4 method for signature 'ModifierSet'
subset(x, name, pos = 1L, ...)

## S4 method for signature 'ModifierSet,GRanges'
subsetByCoord(x, coord, ...)

## S4 method for signature 'ModifierSet,GRangesList'
subsetByCoord(x, coord, ...)

## S4 method for signature 'Modifier,GRanges'
labelByCoord(x, coord, ...)

## S4 method for signature 'Modifier,GRangesList'
labelByCoord(x, coord, ...)

## S4 method for signature 'ModifierSet,GRanges'
labelByCoord(x, coord, ...)

## S4 method for signature 'ModifierSet,GRangesList'

```

```
labelByCoord(x, coord, ...)  
  
## S4 method for signature 'SplitDataFrameList,GRanges'  
subsetByCoord(x, coord, ...)  
  
## S4 method for signature 'SequenceData'  
subset(x, name, pos = 1L, ...)  
  
## S4 method for signature 'SequenceData,GRanges'  
subsetByCoord(x, coord, ...)  
  
## S4 method for signature 'SequenceData,GRangesList'  
subsetByCoord(x, coord, ...)  
  
## S4 method for signature 'SequenceDataSet'  
subset(x, name, pos = 1L, ...)  
  
## S4 method for signature 'SequenceDataSet,GRanges'  
subsetByCoord(x, coord, ...)  
  
## S4 method for signature 'SequenceDataSet,GRangesList'  
subsetByCoord(x, coord, ...)  
  
## S4 method for signature 'SequenceDataList'  
subset(x, name, pos = 1L, ...)  
  
## S4 method for signature 'SequenceDataList,GRanges'  
subsetByCoord(x, coord, ...)  
  
## S4 method for signature 'SequenceDataList,GRangesList'  
subsetByCoord(x, coord, ...)  
  
## S4 method for signature 'SequenceData,GRanges'  
labelByCoord(x, coord, ...)  
  
## S4 method for signature 'SequenceData,GRangesList'  
labelByCoord(x, coord, ...)  
  
## S4 method for signature 'SequenceDataSet,GRanges'  
labelByCoord(x, coord, ...)  
  
## S4 method for signature 'SequenceDataSet,GRangesList'  
labelByCoord(x, coord, ...)  
  
## S4 method for signature 'SequenceDataList,GRanges'  
labelByCoord(x, coord, ...)  
  
## S4 method for signature 'SequenceDataList,GRangesList'  
labelByCoord(x, coord, ...)
```

**Arguments**

x	a <code>SequenceData</code> , <code>SequenceDataSet</code> , <code>SequenceDataList</code> , <code>Modifier</code> or <code>ModifierSet</code> object.
coord	coordinates of position to subset to. Either a <code>GRanges</code> or a <code>GRangesList</code> object. For both types the 'Parent' column is expected to match the transcript name.
...	Optional parameters: <ul style="list-style-type: none"> <li>• <code>type</code>: the modification type used for subsetting. By default this is derived from the <code>modType(x)</code>, but it can be overwritten using <code>type</code>. It must be a valid <code>shortName</code> for a modification according to <code>shortName(ModRNAString())</code> or <code>shortName(ModDNAString())</code> (depending on the type of <code>Modifier</code> class) and of course be present in metadata column <code>mod</code> of <code>coord</code>. To disable subsetting based on type, set <code>type = NA</code>.</li> <li>• <code>flanking</code>: a single integer value to select how many flanking position should be included in the subset (default: <code>flanking = 0L</code>).</li> <li>• <code>merge</code>: <code>TRUE</code> or <code>FALSE</code>: Should the overlapping selections be merged? This is particular important, if <code>flanking</code> value <math>\neq 0L</math> are set. (default: <code>merge = TRUE</code>).</li> <li>• <code>perTranscript</code>: <code>TRUE</code> or <code>FALSE</code>: Should the positions labeled per transcript and not per chromosome? (default: <code>perTranscript = FALSE</code>).</li> </ul>
name	Optional: Limit results to one specific transcript.
pos	Optional: Limit results to a specific position.

**Value**

If 'x' is a

- `SequenceData` or `Modifier`: a `SplitDataFrameList` with elements per transcript.
- `SequenceDataSet`, `SequenceDataList` or `ModifierSet`: a `SimpleList` of `SplitDataFrameList` with elements per transcript.

**Examples**

```
data(msi, package="RNAmodR")
mod <- modifications(msi)
coord <- unique(unlist(mod))
coord$score <- NULL
coord$sd <- NULL
subsetByCoord(msi, coord)
```

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