

An Introduction to *GenomeInfoDb*

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1 Introduction

The *GenomeInfoDb* provides an interface to access seqlevelsStyles (such as UCSC, NCBI, Ensembl) and their supported mappings for organisms. For instance, for Homo sapiens, seqlevelsStyle "UCSC" maps to "chr1", "chr2", ..., "chrX", "chrY". The section below introduces these functions with examples.

2 Functionality for all existing organisms

2.1 genomeStyles

The `genomeStyles` lists out for each organism, the `seqlevelsStyles` and their mappings.

```
seqmap <- genomeStyles()
head(seqmap,n=2)

## $Arabidopsis_thaliana
##   circular auto sex NCBI TAIR9 Ensembl
## 1  FALSE TRUE FALSE 1 Chr1 1
## 2  FALSE TRUE FALSE 2 Chr2 2
## 3  FALSE TRUE FALSE 3 Chr3 3
## 4  FALSE TRUE FALSE 4 Chr4 4
## 5  FALSE TRUE FALSE 5 Chr5 5
## 6   TRUE FALSE FALSE MT ChrM Mt
## 7   TRUE FALSE TRUE Pltd ChrC Pt
##
## $Caenorhabditis_elegans
##   circular auto sex NCBI UCSC Ensembl
## 1  FALSE TRUE FALSE I chrI I
## 2  FALSE TRUE FALSE II chrII II
## 3  FALSE TRUE FALSE III chrIII III
## 4  FALSE TRUE FALSE IV chrIV IV
## 5  FALSE TRUE FALSE V chrV V
## 6  FALSE FALSE TRUE X chrX X
## 7   TRUE TRUE FALSE MT chrM MtDNA
```

Organism's supported by GenomeInfoDb can be found by :

```
names(genomeStyles())

## [1] "Arabidopsis_thaliana" "Caenorhabditis_elegans"
## [3] "Canis_familiaris" "Cyanidioschyzon_merolae"
## [5] "Drosophila_melanogaster" "Gossypium_hirsutum"
## [7] "Homo_sapiens" "Mus_musculus"
## [9] "Oryza_sativa" "Populus_trichocarpa"
## [11] "Rattus_norvegicus" "Saccharomyces_cerevisiae"
## [13] "Zea_mays"
```

If one knows the organism one is interested in, then we can directly access the information for the given organism along. Each function accepts an argument called `species` which as "genus species", the default is "Homo sapiens". In the following example we list out only the first five entries returned by the code snippet.

```
head(genomeStyles("Homo_sapiens"),5)

##   circular auto sex NCBI UCSC dbSNP Ensembl
## 1  FALSE TRUE FALSE 1 chr1 ch1 1
## 2  FALSE TRUE FALSE 2 chr2 ch2 2
## 3  FALSE TRUE FALSE 3 chr3 ch3 3
## 4  FALSE TRUE FALSE 4 chr4 ch4 4
## 5  FALSE TRUE FALSE 5 chr5 ch5 5
```

We can also check if a given style is supported by GenomeInfoDb for a given species. For example, if we want to know if "UCSC" mapping is supported for "Homo sapiens" we can ask :

```
"UCSC" %in% names(genomeStyles("Homo_sapiens"))  
## [1] TRUE
```

2.2 extractSeqlevels

We can also extract the desired seqlevelsStyle from a given organism using the `extractSeqlevels`

```
extractSeqlevels(species="Arabidopsis_thaliana", style="NCBI")  
## [1] "1" "2" "3" "4" "5" "MT" "Pltd"
```

2.3 extractSeqlevelsByGroup

We can also extract the desired seqlevelsStyle from a given organism based on a group (Group - 'auto' denotes autosomes, 'circular' denotes circular chromosomes and 'sex' denotes sex chromosomes; the default is all chromosomes are returned).

```
extractSeqlevelsByGroup(species="Arabidopsis_thaliana", style="NCBI",  
                        group="auto")  
## [1] "1" "2" "3" "4" "5"
```

2.4 seqlevelsStyle

We can find the seqname Style for a given character vector by using the `seqlevelsStyle`

```
seqlevelsStyle(paste0("chr", c(1:30)))  
## [1] "UCSC"  
seqlevelsStyle(c("2L", "2R", "X", "Xhet"))  
## [1] "NCBI"
```

2.5 seqlevelsInGroup

We can also subset a given character vector containing seqnames using the `seqlevelsInGroup`. We currently support 3 groups: 'auto' for autosomes, 'sex' for allosomes/sex chromosomes and circular for 'circular' chromosomes. The user can also provide the style and species they are working with. In the following examples, we extract the sex, auto and circular chromosomes for Homo sapiens :

```
newchr <- paste0("chr", c(1:22, "X", "Y", "M", "1_gl000192_random", "4_ctg9_hap1"))  
seqlevelsInGroup(newchr, group="sex")
```

```
## [1] "chrX" "chrY"
seqlevelsInGroup(newchr, group="auto")
## [1] "chr1" "chr2" "chr3" "chr4" "chr5" "chr6" "chr7" "chr8" "chr9"
## [10] "chr10" "chr11" "chr12" "chr13" "chr14" "chr15" "chr16" "chr17" "chr18"
## [19] "chr19" "chr20" "chr21" "chr22"
seqlevelsInGroup(newchr, group="circular")
## [1] "chrM"
seqlevelsInGroup(newchr, group="sex", "Homo_sapiens", "UCSC")
## [1] "chrX" "chrY"
```

if we have a vector containing seqnames and we want to verify the species and style for them, we can use:

```
seqnames <- c("chr1", "chr9", "chr2", "chr3", "chr10")
all(seqnames %in% extractSeqlevels("Homo_sapiens", "UCSC"))
## [1] TRUE
```

2.6 orderSeqlevels

The `orderSeqlevels` can return the order of a given character vector which contains seqnames. In the following example, we show how you can find the order for a given seqnames character vector.

```
seqnames <- c("chr1", "chr9", "chr2", "chr3", "chr10")
orderSeqlevels(seqnames)
## [1] 1 3 4 2 5
seqnames[orderSeqlevels(seqnames)]
## [1] "chr1" "chr2" "chr3" "chr9" "chr10"
```

2.7 rankSeqlevels

The `rankSeqlevels` can return the rank of a given character vector which contains seqnames. In the following example, we show how you can find the rank for a given seqnames character vector.

```
seqnames <- c("chr1", "chr9", "chr2", "chr3", "chr10")
rankSeqlevels(seqnames)
## [1] 1 4 2 3 5
```

2.8 mapSeqlevels

Returns a matrix with 1 column per supplied sequence name and 1 row per sequence renaming map compatible with the specified style. If `best.only` is `TRUE` (the default), only the "best" renaming maps (i.e. the rows with less NAs) are returned.

```
mapSeqlevels(c("chrII", "chrIII", "chrM"), "NCBI")

## chrII chrIII chrM
## "II" "III" "MT"
```

We also have several seqlevel utility functions. Let us construct a basic GRanges and show how these functions can be used. .

```
gr <- GRanges(paste0("ch", 1:35), IRanges(1:35, width=5))
gr

## GRanges object with 35 ranges and 0 metadata columns:
##      seqnames      ranges strand
##      <Rle> <IRanges> <Rle>
##   [1]    ch1        1-5      *
##   [2]    ch2        2-6      *
##   [3]    ch3        3-7      *
##   [4]    ch4        4-8      *
##   [5]    ch5        5-9      *
##   ...      ...          ...    ...
##  [31]   ch31       31-35      *
##  [32]   ch32       32-36      *
##  [33]   ch33       33-37      *
##  [34]   ch34       34-38      *
##  [35]   ch35       35-39      *
## -----
## seqinfo: 35 sequences from an unspecified genome; no seqlengths
```

As you can see, we have "ch" instead of "chr" for chromosome names. We can use `renameSeqlevels` to change the "ch" to "chr"

2.9 renameSeqlevels

As the first argument - it takes the object whose seqlevels we need to change, and as the second argument it takes a named vector which has the changes.

```
newnames <- paste0("chr", 1:35)
names(newnames) <- paste0("ch", 1:35)
head(newnames)

##   ch1   ch2   ch3   ch4   ch5   ch6
## "chr1" "chr2" "chr3" "chr4" "chr5" "chr6"

gr <- renameSeqlevels(gr, newnames)
gr

## GRanges object with 35 ranges and 0 metadata columns:
##      seqnames      ranges strand
##      <Rle> <IRanges> <Rle>
##   [1]   chr1        1-5      *
##   [2]   chr2        2-6      *
##   [3]   chr3        3-7      *
##   [4]   chr4        4-8      *
##   [5]   chr5        5-9      *
```

```
##      ...      ...      ...      ...
## [31] chr31      31-35      *
## [32] chr32      32-36      *
## [33] chr33      33-37      *
## [34] chr34      34-38      *
## [35] chr35      35-39      *
## -----
## seqinfo: 35 sequences from an unspecified genome; no seqlengths
```

Humans have just 22 primary chromosomes - but here we have some extra seqlevels which we want to remove - there are several ways we can achieve this:

2.10 dropSeqlevels

Here the second argument is the seqlevels that you want to drop. Because these seqlevels are in use (i.e. have ranges on them), the ranges on these sequences need to be removed before the seqlevels can be dropped. We call this *pruning*. The `pruning.mode` argument controls how to prune `gr`. Unlike for list-like objects (e.g. `GRangesList`) for which pruning can be done in various ways, pruning a `GRanges` object is straightforward and achieved by specifying `pruning.mode="coarse"`.

```
dropSeqlevels(gr, paste0("chr",23:35), pruning.mode="coarse")

## GRanges object with 22 ranges and 0 metadata columns:
##      seqnames      ranges strand
##      <Rle> <IRanges> <Rle>
## [1] chr1      1-5      *
## [2] chr2      2-6      *
## [3] chr3      3-7      *
## [4] chr4      4-8      *
## [5] chr5      5-9      *
## ...      ...      ...      ...
## [18] chr18     18-22     *
## [19] chr19     19-23     *
## [20] chr20     20-24     *
## [21] chr21     21-25     *
## [22] chr22     22-26     *
## -----
## seqinfo: 22 sequences from an unspecified genome; no seqlengths
```

2.11 keepSeqlevels

Here the second argument is the seqlevels that you want to keep.

```
keepSeqlevels(gr, paste0("chr",1:22), pruning.mode="coarse")

## GRanges object with 22 ranges and 0 metadata columns:
##      seqnames      ranges strand
##      <Rle> <IRanges> <Rle>
## [1] chr1      1-5      *
## [2] chr2      2-6      *
```

```
##      [3]      chr3      3-7      *
##      [4]      chr4      4-8      *
##      [5]      chr5      5-9      *
##      ...      ...      ...      ...
##     [18]     chr18     18-22     *
##     [19]     chr19     19-23     *
##     [20]     chr20     20-24     *
##     [21]     chr21     21-25     *
##     [22]     chr22     22-26     *
##     -----
##     seqinfo: 22 sequences from an unspecified genome; no seqlengths
```

2.12 keepStandardChromosomes

This function internally uses the pre-defined tables inside *GenomeInfoDb* to find the correct seqlevels according to the sequence style of the object.

```
keepStandardChromosomes(gr, pruning.mode="coarse")

## GRanges object with 35 ranges and 0 metadata columns:
##      seqnames      ranges strand
##      <Rle> <IRanges> <Rle>
##      [1]      chr1      1-5      *
##      [2]      chr2      2-6      *
##      [3]      chr3      3-7      *
##      [4]      chr4      4-8      *
##      [5]      chr5      5-9      *
##      ...      ...      ...      ...
##     [31]     chr31     31-35     *
##     [32]     chr32     32-36     *
##     [33]     chr33     33-37     *
##     [34]     chr34     34-38     *
##     [35]     chr35     35-39     *
##     -----
##     seqinfo: 35 sequences from an unspecified genome; no seqlengths
```

One can also specify the optional species argument to be more precise.

```
plantgr <- GRanges(c(1:5,"MT","Pltd"), IRanges(1:7,width=5))
keepStandardChromosomes(plantgr, species="Arabidopsis thaliana",
                        pruning.mode="coarse")

## GRanges object with 7 ranges and 0 metadata columns:
##      seqnames      ranges strand
##      <Rle> <IRanges> <Rle>
##      [1]         1      1-5      *
##      [2]         2      2-6      *
##      [3]         3      3-7      *
##      [4]         4      4-8      *
##      [5]         5      5-9      *
##      [6]        MT      6-10     *
##      [7]       Pltd      7-11     *
```

```
## -----
## seqinfo: 7 sequences from an unspecified genome; no seqlengths
```

3 Seqinfo objects

```
## Note that all the arguments (except 'genome') must have the
## same length. 'genome' can be of length 1, whatever the lengths
## of the other arguments are.
x <- Seqinfo(seqnames=c("chr1", "chr2", "chr3", "chrM"),
             seqlengths=c(100, 200, NA, 15),
             isCircular=c(NA, FALSE, FALSE, TRUE),
             genome="toy")

length(x)
## [1] 4

seqnames(x)
## [1] "chr1" "chr2" "chr3" "chrM"

names(x)
## [1] "chr1" "chr2" "chr3" "chrM"

seqlevels(x)
## [1] "chr1" "chr2" "chr3" "chrM"

seqlengths(x)
## chr1 chr2 chr3 chrM
## 100 200 NA 15

isCircular(x)
## chr1 chr2 chr3 chrM
## NA FALSE FALSE TRUE

genome(x)
## chr1 chr2 chr3 chrM
## "toy" "toy" "toy" "toy"

x[c("chrY", "chr3", "chr1")] # subset by names

## Seqinfo object with 3 sequences from 2 genomes (NA, toy):
## seqnames seqlengths isCircular genome
## chrY      NA      NA <NA>
## chr3      NA      FALSE toy
## chr1      100     NA toy

## Rename, drop, add and/or reorder the sequence levels:
xx <- x
seqlevels(xx) <- sub("chr", "ch", seqlevels(xx)) # rename
xx

## Seqinfo object with 4 sequences (1 circular) from toy genome:
## seqnames seqlengths isCircular genome
```



```
##      ch1          100          NA      toy
##      ch2          200         FALSE    toy
##      ch3           NA         FALSE    toy
##      chM           15          TRUE     toy

seqlevels(xx) <- rev(seqlevels(xx)) # reorder
xx

## Seqinfo object with 4 sequences (1 circular) from toy genome:
##      seqnames seqlengths isCircular genome
##      chM          15          TRUE     toy
##      ch3           NA         FALSE    toy
##      ch2          200         FALSE    toy
##      ch1          100          NA      toy

seqlevels(xx) <- c("ch1", "ch2", "chY") # drop/add/reorder
xx

## Seqinfo object with 3 sequences from 2 genomes (toy, NA):
##      seqnames seqlengths isCircular genome
##      ch1          100          NA      toy
##      ch2          200         FALSE    toy
##      chY           NA          NA     <NA>

seqlevels(xx) <- c(chY="Y", ch1="1", "22") # rename/reorder/drop/add
xx

## Seqinfo object with 3 sequences from 2 genomes (NA, toy):
##      seqnames seqlengths isCircular genome
##      Y           NA          NA     <NA>
##      1           100          NA      toy
##      22          NA          NA     <NA>

y <- Seqinfo(seqnames=c("chr3", "chr4", "chrM"),
              seqlengths=c(300, NA, 15))
y

## Seqinfo object with 3 sequences from an unspecified genome:
##      seqnames seqlengths isCircular genome
##      chr3      300          NA     <NA>
##      chr4      NA          NA     <NA>
##      chrM      15          NA     <NA>

merge(x, y) # rows for chr3 and chrM are merged

## Warning in .merge_two_Seqinfo_objects(x, y): Each of the 2 combined objects
has sequence levels not in the other:
## - in 'x': chr1, chr2
## - in 'y': chr4
## Make sure to always combine/compare objects based on the same reference
## genome (use suppressWarnings() to suppress this warning).

## Seqinfo object with 5 sequences (1 circular) from 2 genomes (toy, NA):
##      seqnames seqlengths isCircular genome
##      chr1      100          NA      toy
##      chr2      200         FALSE    toy
```

```
## chr3      300      FALSE    toy
## chrM      15       TRUE     toy
## chr4      NA       NA      <NA>

suppressWarnings(merge(x, y))

## Seqinfo object with 5 sequences (1 circular) from 2 genomes (toy, NA):
## seqnames seqlengths isCircular genome
## chr1      100       NA      toy
## chr2      200      FALSE    toy
## chr3      300      FALSE    toy
## chrM      15       TRUE     toy
## chr4      NA       NA      <NA>

## Note that, strictly speaking, merging 2 Seqinfo objects is not
## a commutative operation, i.e., in general 'z1 <- merge(x, y)'
## is not identical to 'z2 <- merge(y, x)'. However 'z1' and 'z2'
## are guaranteed to contain the same information (i.e. the same
## rows, but typically not in the same order):
suppressWarnings(merge(y, x))

## Seqinfo object with 5 sequences (1 circular) from 2 genomes (toy, NA):
## seqnames seqlengths isCircular genome
## chr3      300      FALSE    toy
## chr4      NA       NA      <NA>
## chrM      15       TRUE     toy
## chr1      100       NA      toy
## chr2      200      FALSE    toy

## This contradicts what 'x' says about circularity of chr3 and chrM:
isCircular(y)[c("chr3", "chrM")] <- c(TRUE, FALSE)
y

## Seqinfo object with 3 sequences (1 circular) from an unspecified genome:
## seqnames seqlengths isCircular genome
## chr3      300       TRUE     <NA>
## chr4      NA       NA      <NA>
## chrM      15      FALSE     <NA>

if (interactive()) {
  merge(x, y) # raises an error
}
```

4 Examples

4.1 converting seqlevel styles (eg:UCSC to NCBI)

A quick example using *Drosophila Melanogaster*. The txdb object contains seqlevels in UCSC style, we want to convert them to NCBI

```
txdb <- TxDb.Dmelanogaster.UCSC.dm3.ensGene
seqlevels(txdb)
```

```
## [1] "chr2L"      "chr2R"      "chr3L"      "chr3R"      "chr4"      "chrX"
## [7] "chrU"       "chrM"       "chr2LHet"   "chr2RHet"   "chr3LHet"   "chr3RHet"
## [13] "chrXHet"    "chrYHet"    "chrUextra"
```

genomeStyles("Drosophila melanogaster")

	circular	sex	auto	NCBI	UCSC	Ensembl
## 1	FALSE	FALSE	TRUE	2L	chr2L	2L
## 2	FALSE	FALSE	TRUE	2R	chr2R	2R
## 3	FALSE	FALSE	TRUE	3L	chr3L	3L
## 4	FALSE	FALSE	TRUE	3R	chr3R	3R
## 5	FALSE	FALSE	TRUE	4	chr4	4
## 6	FALSE	TRUE	FALSE	X	chrX	X
## 7	FALSE	TRUE	FALSE	Y	chrY	Y
## 8	TRUE	FALSE	FALSE	MT	chrM	dmel_mitochondrion_genome
## 9	FALSE	FALSE	FALSE	2LHet	chr2LHet	2LHet
## 10	FALSE	FALSE	FALSE	2Rhet	chr2RHet	2RHet
## 11	FALSE	FALSE	FALSE	3LHet	chr3LHet	3LHet
## 12	FALSE	FALSE	FALSE	3RHet	chr3RHet	3RHet
## 13	FALSE	FALSE	FALSE	Xhet	chrXHet	XHet
## 14	FALSE	FALSE	FALSE	Yhet	chrYHet	YHet
## 15	FALSE	FALSE	FALSE	Un	chrU	U
## 16	FALSE	FALSE	FALSE	<NA>	chrUextra	Uextra

mapSeqlevels(seqlevels(txdb), "NCBI")

	chr2L	chr2R	chr3L	chr3R	chr4	chrX	chrU
##	"2L"	"2R"	"3L"	"3R"	"4"	"X"	"Un"
##	chrM	chr2LHet	chr2RHet	chr3LHet	chr3RHet	chrXHet	chrYHet
##	"MT"	"2LHet"	"2Rhet"	"3LHet"	"3RHet"	"Xhet"	"Yhet"
##	chrUextra						
##	NA						

4.2 converting styles and removing unwanted seqlevels

Suppose we read in a Bam file or a BED file and the resulting GRanges have a lot of seqlevels which are not required by your analysis or you want to rename the seqlevels from the current style to your own style (eg: UCSC to NCBI), we can use the functionality provided by GenomeInfoDb to do that.

Let us say that we have extracted the seqlevels of the Seqinfo object (say GRanges from a BED file) in a variable called "sequence".

```
sequence <- seqlevels(x)

## sequence is in UCSC format and we want NCBI style
newStyle <- mapSeqlevels(sequence, "NCBI")
newStyle <- newStyle[complete.cases(newStyle)] # removing NA cases.

## rename the seqlevels
x <- renameSeqlevels(x, newStyle)

## keep only the seqlevels you want (say autosomes)
```

```
auto <- extractSeqlevelsByGroup(species="Homo sapiens", style="NCBI",  
                                group="auto")  
x <- keepSeqlevels(x, auto)
```

5 Session Information

Here is the output of `sessionInfo` on the system on which this document was compiled:

```
toLatex(sessionInfo())
```

- R version 4.3.2 Patched (2023-11-13 r85521), x86_64-pc-linux-gnu
- Locale: LC_CTYPE=en_US.UTF-8, LC_NUMERIC=C, LC_TIME=en_GB, LC_COLLATE=C, LC_MONETARY=en_US.UTF-8, LC_MESSAGES=en_US.UTF-8, LC_PAPER=en_US.UTF-8, LC_NAME=C, LC_ADDRESS=C, LC_TELEPHONE=C, LC_MEASUREMENT=en_US.UTF-8, LC_IDENTIFICATION=C
- Time zone: America/New_York
- TZcode source: system (glibc)
- Running under: Ubuntu 22.04.4 LTS
- Matrix products: default
- BLAS: /home/biocbuild/bbs-3.18-bioc/R/lib/libRblas.so
- LAPACK: /usr/lib/x86_64-linux-gnu/lapack/liblapack.so.3.10.0
- Base packages: base, datasets, grDevices, graphics, methods, stats, stats4, utils
- Other packages: AnnotationDbi 1.64.1, Biobase 2.62.0, BiocGenerics 0.48.1, BiocStyle 2.30.0, GenomeInfoDb 1.38.7, GenomicFeatures 1.54.3, GenomicRanges 1.54.1, IRanges 2.36.0, S4Vectors 0.40.2, TxDb.Dmelanogaster.UCSC.dm3.ensGene 3.2.2
- Loaded via a namespace (and not attached): BiocFileCache 2.10.1, BiocIO 1.12.0, BiocManager 1.30.22, BiocParallel 1.36.0, Biostrings 2.70.2, DBI 1.2.2, DelayedArray 0.28.0, GenomeInfoDbData 1.2.11, GenomicAlignments 1.38.2, KEGGREST 1.42.0, Matrix 1.6-5, MatrixGenerics 1.14.0, R6 2.5.1, RCurl 1.98-1.14, RSQLite 2.3.5, Rsamtools 2.18.0, S4Arrays 1.2.1, SparseArray 1.2.4, SummarizedExperiment 1.32.0, XML 3.99-0.16.1, XVector 0.42.0, abind 1.4-5, biomaRt 2.58.2, bit 4.0.5, bit64 4.0.5, bitops 1.0-7, blob 1.2.4, bookdown 0.38, bslib 0.6.1, cachem 1.0.8, cli 3.6.2, codetools 0.2-19, compiler 4.3.2, crayon 1.5.2, curl 5.2.1, dbplyr 2.4.0, digest 0.6.34, dplyr 1.1.4, evaluate 0.23, fansi 1.0.6, fastmap 1.1.1, filelock 1.0.3, generics 0.1.3, glue 1.7.0, grid 4.3.2, highr 0.10, hms 1.1.3, htmltools 0.5.7, httr 1.4.7, jquerylib 0.1.4, jsonlite 1.8.8, knitr 1.45, lattice 0.22-5, lifecycle 1.0.4, magrittr 2.0.3, matrixStats 1.2.0, memoise 2.0.1, parallel 4.3.2, pillar 1.9.0, pkgconfig 2.0.3, png 0.1-8, prettyunits 1.2.0, progress 1.2.3, rappdirs 0.3.3, restfulr 0.0.15, rjson 0.2.21, rlang 1.1.3, rmarkdown 2.26, rtracklayer 1.62.0, sass 0.4.8, stringi 1.8.3, stringr 1.5.1, tibble 3.2.1, tidyselect 1.2.0, tools 4.3.2, utf8 1.2.4, vctrs 0.6.5, xfun 0.42, xml2 1.3.6, yaml 2.3.8, zlibbioc 1.48.0